



Advancing MPN Genomic Profiling with an Amplicon-Based Myeloid NGS Panel

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Learning Objectives:

1

Gain insight into the molecular profiling of MPN including classification and biomarkers implicated in the diagnosis, prognosis, and therapeutic selection for MPN patients.

2

Assess traditional sequential single genes testing via PCR in identifying driver genes.

3

Assess clinical scenarios where an amplicon-based myeloid NGS panel identified critical co-mutations of clinical significance.

4

Explore a proposed diagnostic algorithm that integrates both PCR and NGS for detecting very low variant allele frequencies (VAF) in driver genes.

Myeloproliferative Neoplasms (MPNs):

Hematological disorders, characterized by clonal overproduction of mature WBCs, platelets and/or erythrocytes in the bone marrow and blood

- No evidence of myelodysplasia or dysplastic features on morphology
- Insidious onset, asymptomatic, identified on routine blood count showing increased counts and/or organomegaly
- Progression to myeloid blast phase and secondary AML

Myeloproliferative neoplasms

Myeloproliferative neoplasms

Myeloproliferative neoplasms: Introduction

Chronic myeloid leukaemia

Chronic neutrophilic leukaemia

Chronic eosinophilic leukaemia

Polycythaemia vera

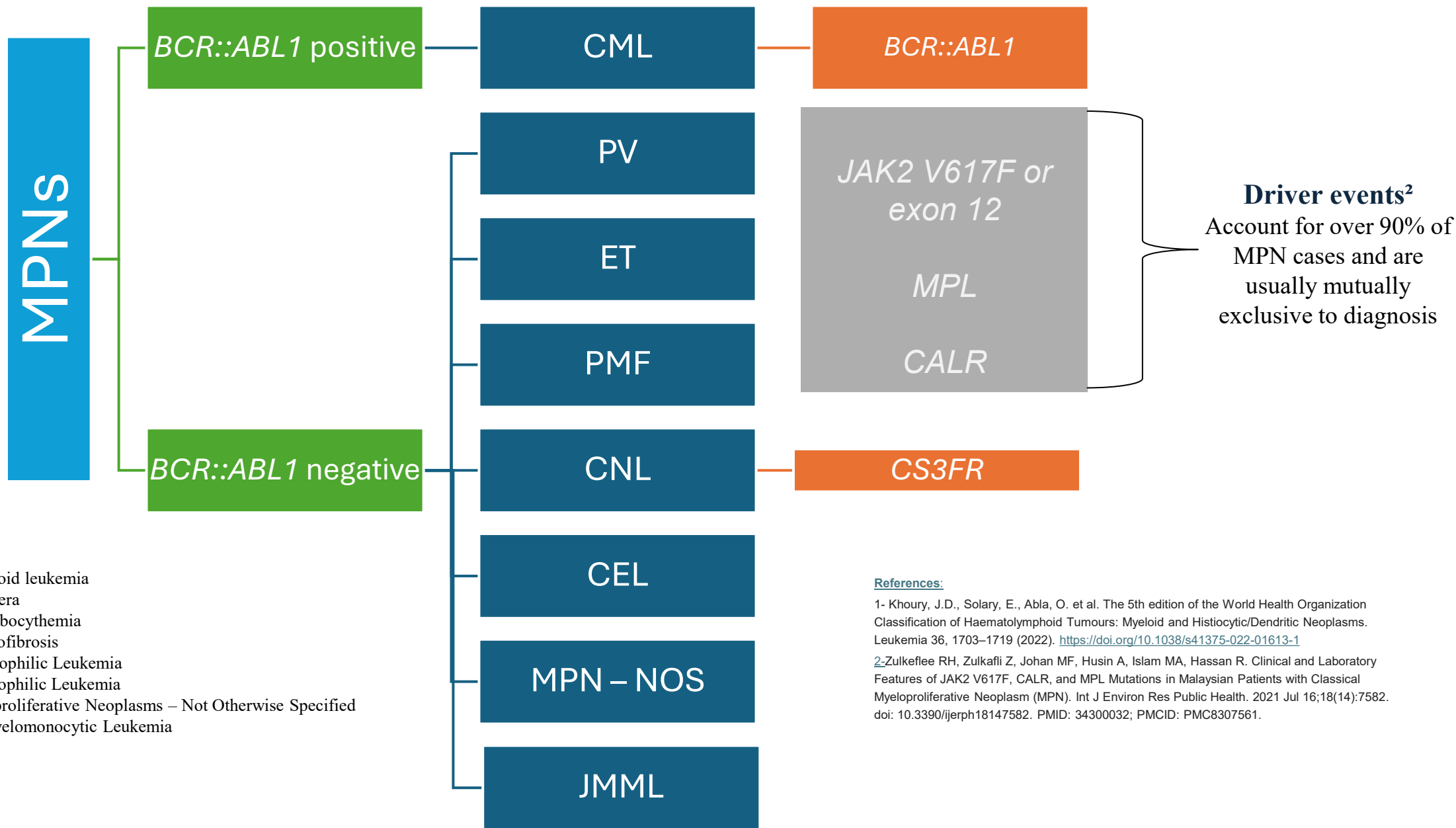
Essential thrombocythaemia

Primary myelofibrosis

Juvenile myelomonocytic leukaemia

Myeloproliferative neoplasm NOS (unclassifiable)

WHO 2022 Classification of MPNs



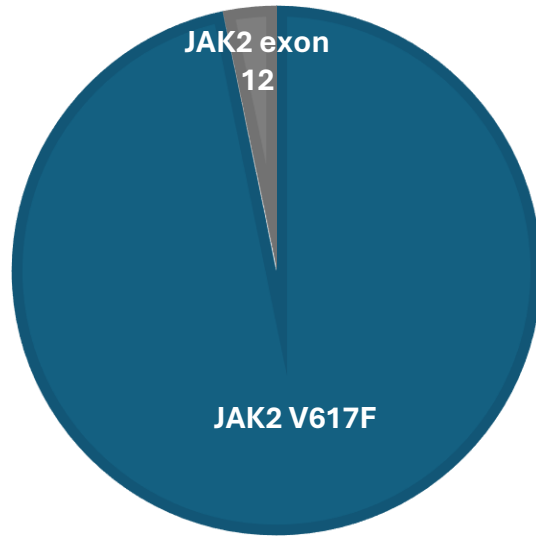
CML: Chronic myeloid leukemia
PV: Polycythemia Vera
ET: Essential Thrombocythemia
PMF: Primary Myelofibrosis
CNL: Chronic Neutrophilic Leukemia
CEL: Chronic Eosinophilic Leukemia
MPN- NOS: Myeloproliferative Neoplasms – Not Otherwise Specified
JMML: Juvenile Myelomonocytic Leukemia

References:

1- Khoury, J.D., Solary, E., Abla, O. et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Myeloid and Histiocytic/Dendritic Neoplasms. *Leukemia* 36, 1703–1719 (2022). <https://doi.org/10.1038/s41375-022-01613-1>
 2-Zulkeflee RH, Zulkafli Z, Johan MF, Husin A, Islam MA, Hassan R. Clinical and Laboratory Features of JAK2 V617F, CALR, and MPL Mutations in Malaysian Patients with Classical Myeloproliferative Neoplasm (MPN). *Int J Environ Res Public Health*. 2021 Jul 16;18(14):7582. doi: 10.3390/ijerph18147582. PMID: 34300032; PMCID: PMC8307561.

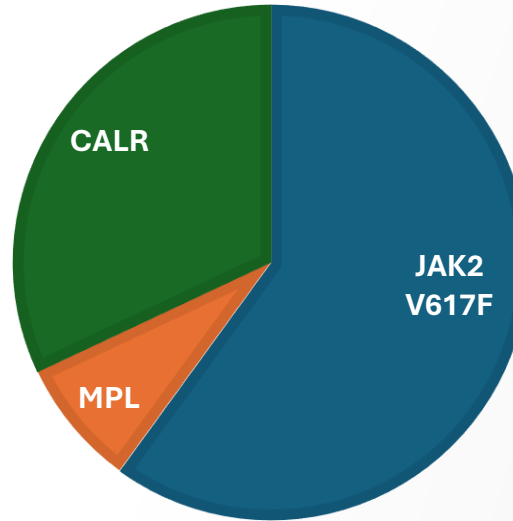
Driver Events in MPNs

☐ ~90% of MPN cases harbor mutations in the three driver genes



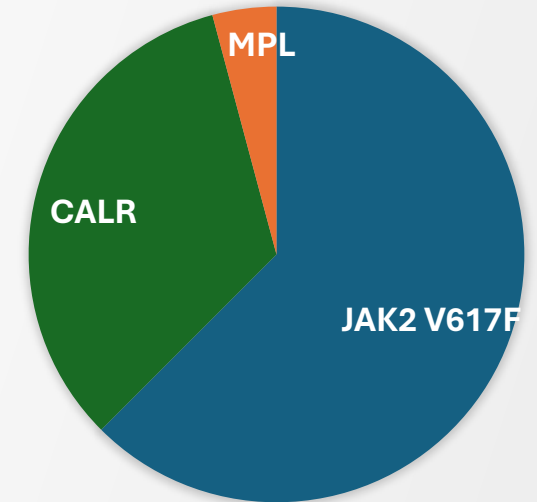
Polycythemia Vera (PV)

- **JAK2 V617F** accounts for ~95% of PV cases
- **JAK2 exon 12** rare insertions and deletions are found in 2% to 3% of patients with PV



Primary Myelofibrosis (PMF)

- **JAK2 V617F** accounts for 60% of MF cases.
- **CALR** frameshift mutations in exon 9 are found in 20% to 30% of all patients with MF
- **MPL W515L/K** accounts for 7-10% of MF cases



Essential Thrombocythemia (ET)

- **JAK2 V617F** accounts for 50-60% of ET cases.
- **CALR** frameshift mutations in exon 9 are found in 20% to 25% of all patients with ET
- **MPL W515L/K** accounts for 5-7% of ET cases

☐ ~10% of MPNs (ET/PMF) are considered “triple negative”: *ASXL1*, *EZH2*, *TET2*, *IDH1*, *IDH2*, *SRSF2*, *SF3B1* mutations

MPN Workup

Diagnosis

Determine diagnosis & subclassification (PV, ET, PMF) via clinical features, blood smear, BM morphology, cytogenetics, and molecular testing (e.g., *JAK2*, *CALR*, and *MPL*)

PCR based sequential testing

Prognosis

Identify other MPN-associated genetic aberrations such as mutations in *ASXL1*, *EZH2*, *TET2*, *IDH1*, *IDH2*, *SRSF2* and *SF3B1* for risk stratification and disease prognostication

Myeloid NGS panel

Therapy Selection

Select appropriate therapy based on diagnosis and risk category such as JAK2 inhibitors

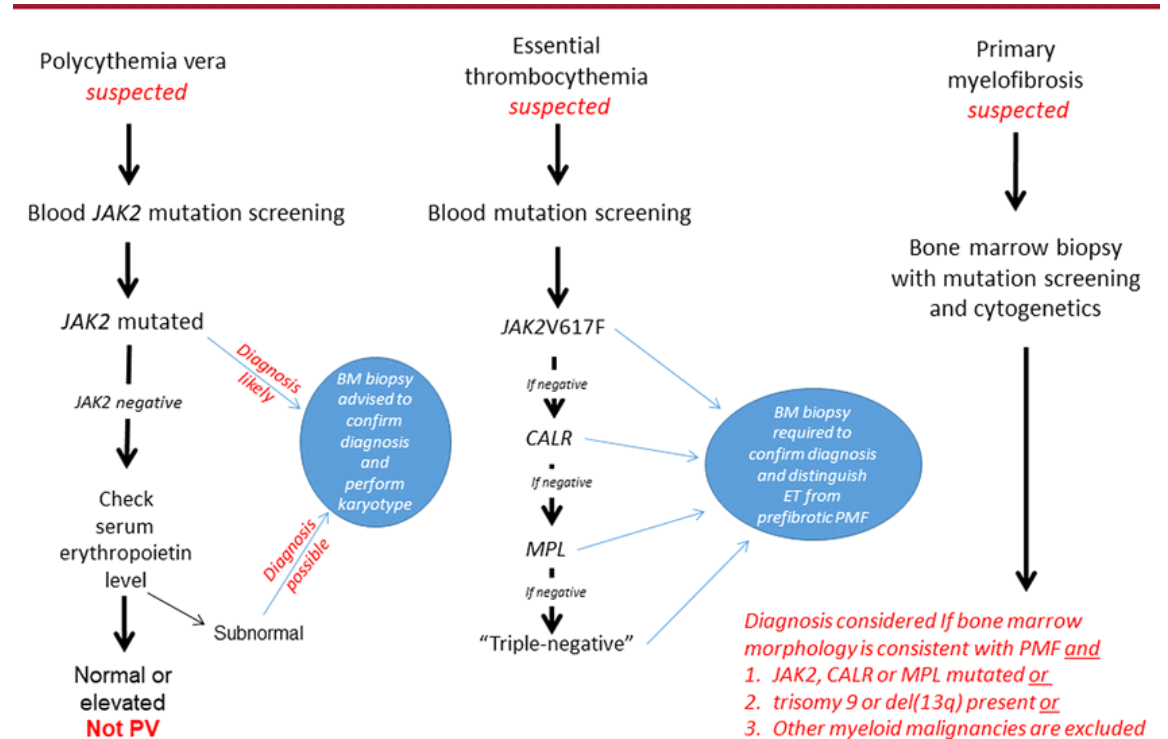
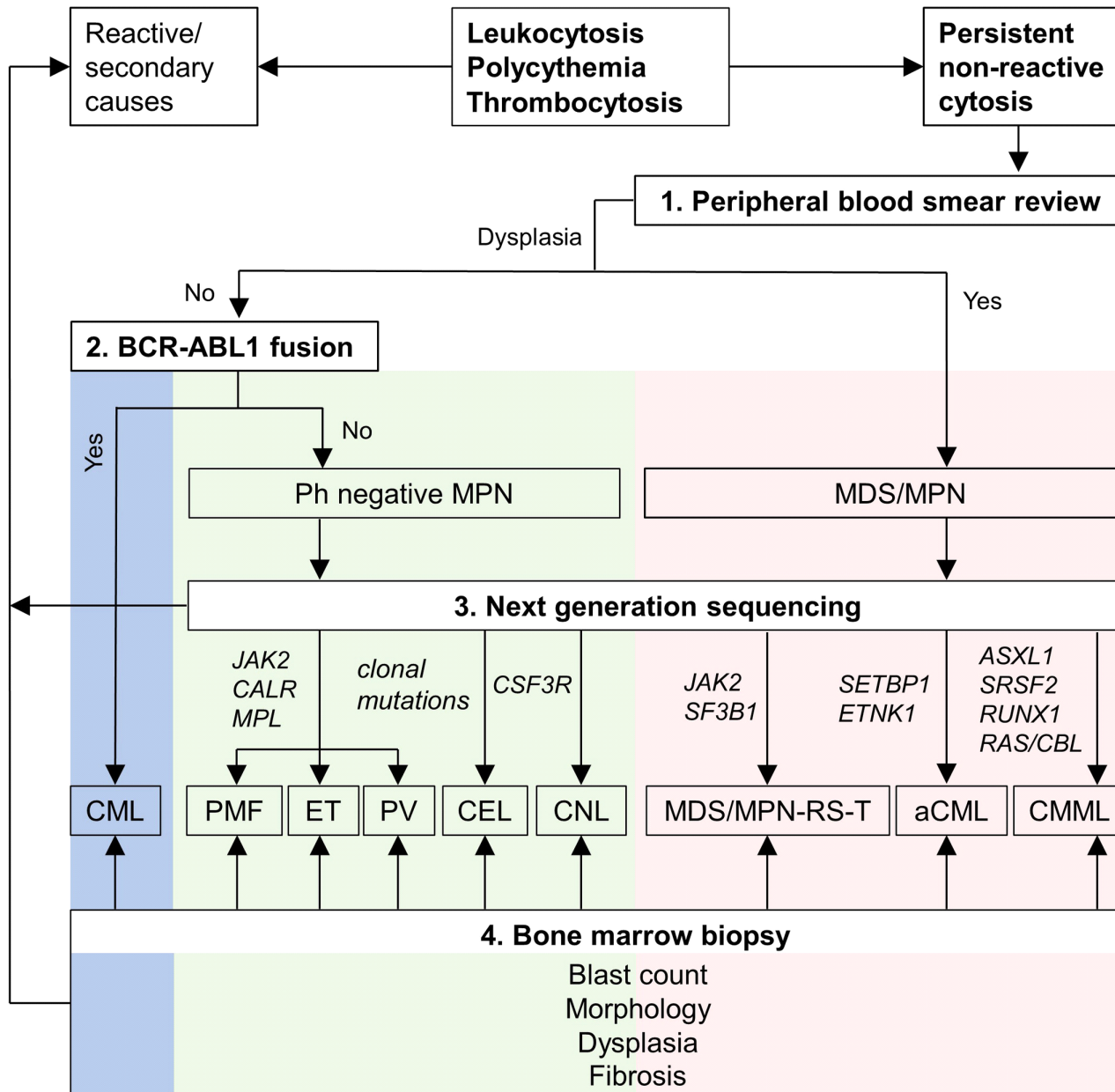
Myeloid NGS panel

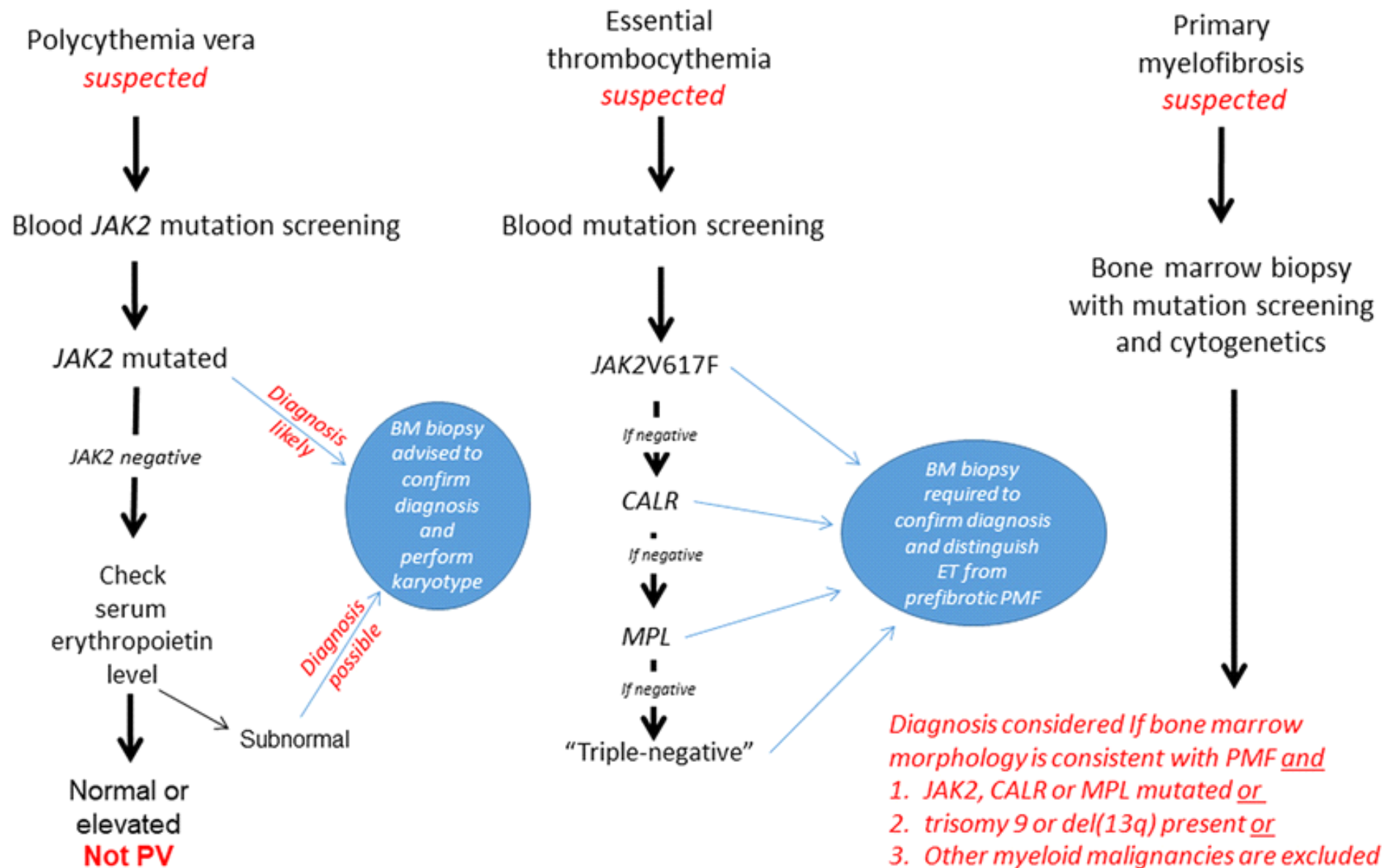
Disease Monitoring (MRD)

Acquired mutations in *JAK2*, *MPL*, *CALR*. Reduction of mutation allele burden are reported in many therapies such as JAK2 inhibitors, allogeneic stem cell transplant, and interferon.

qPCR

Myeloproliferative Neoplasms





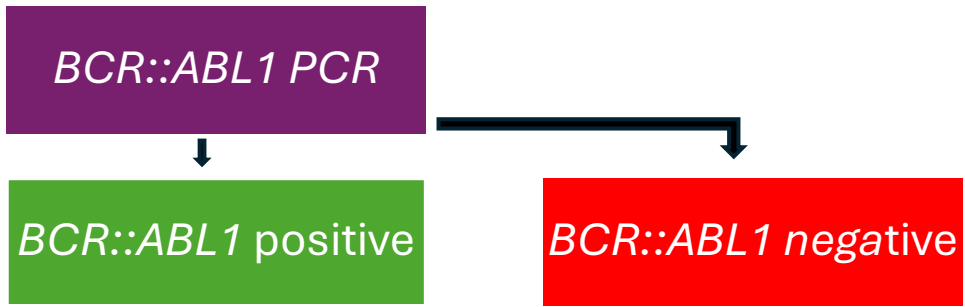
Sequential PCR based testing strategy

BCR::ABL1 PCR

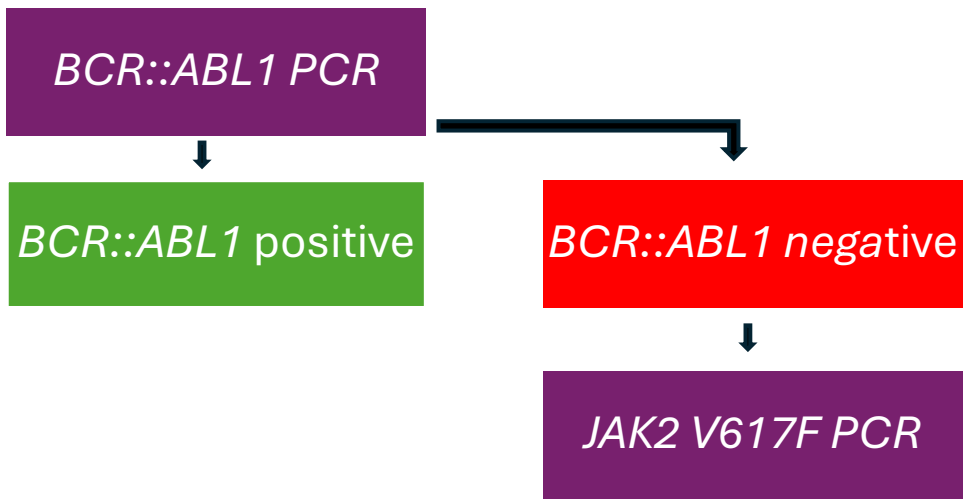


BCR::ABL1 positive

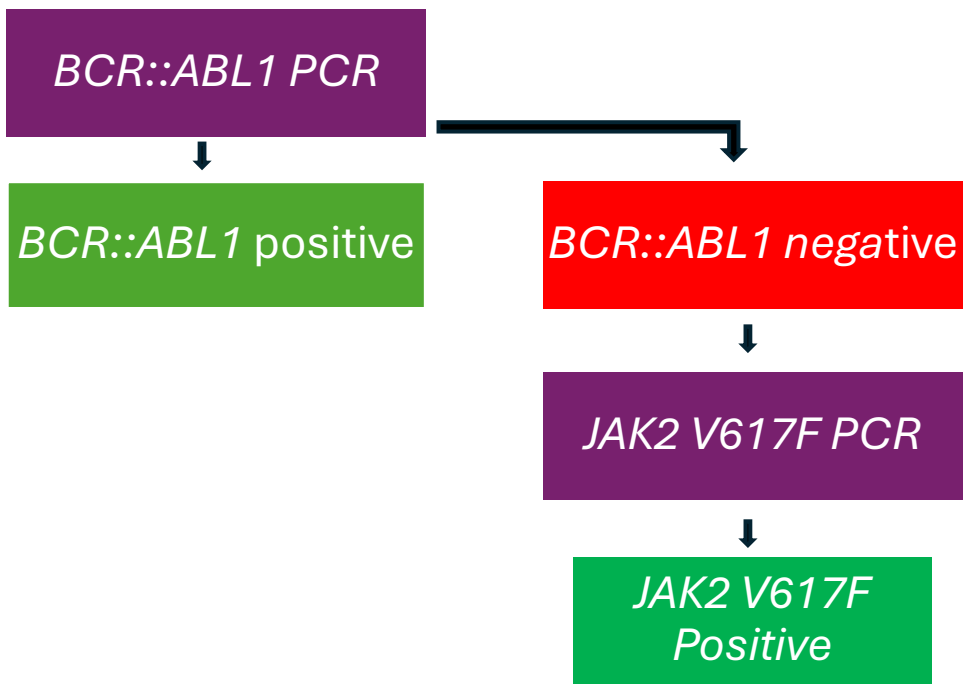
Sequential PCR based testing strategy



Sequential PCR based testing strategy

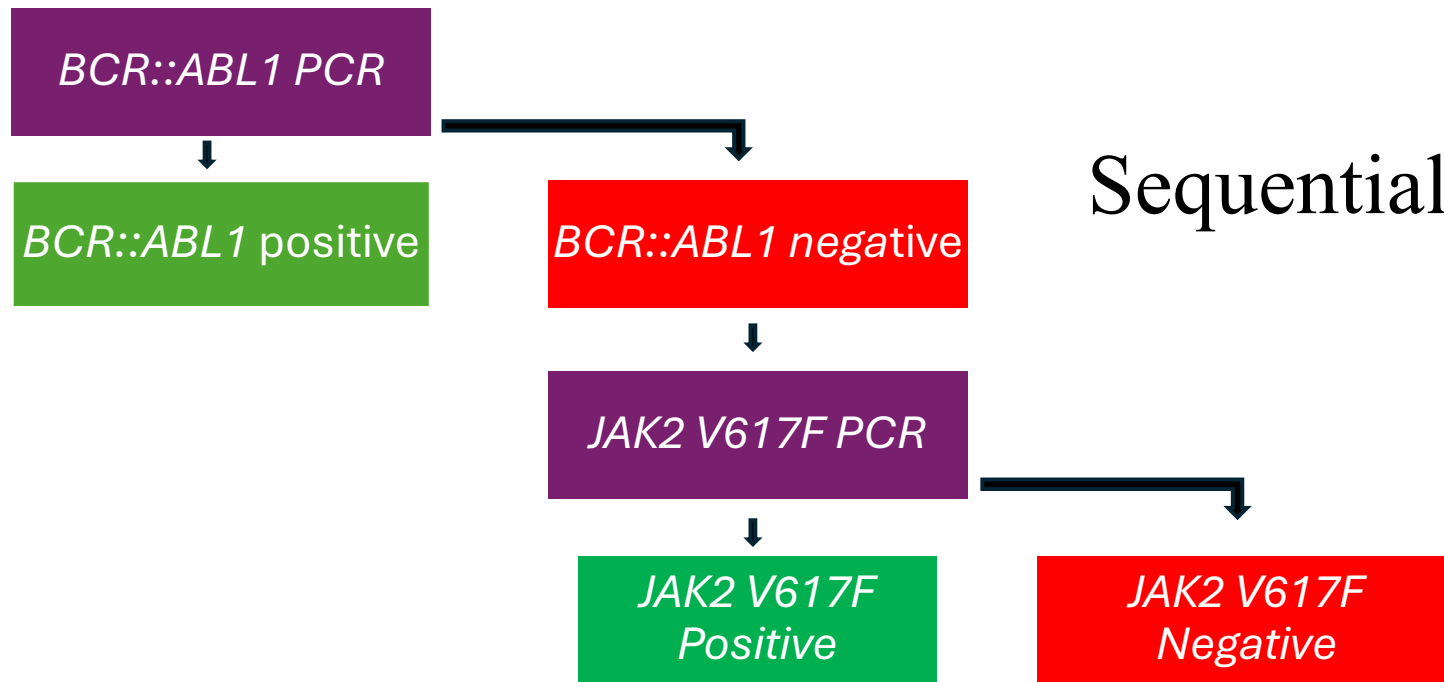


Sequential PCR based testing strategy

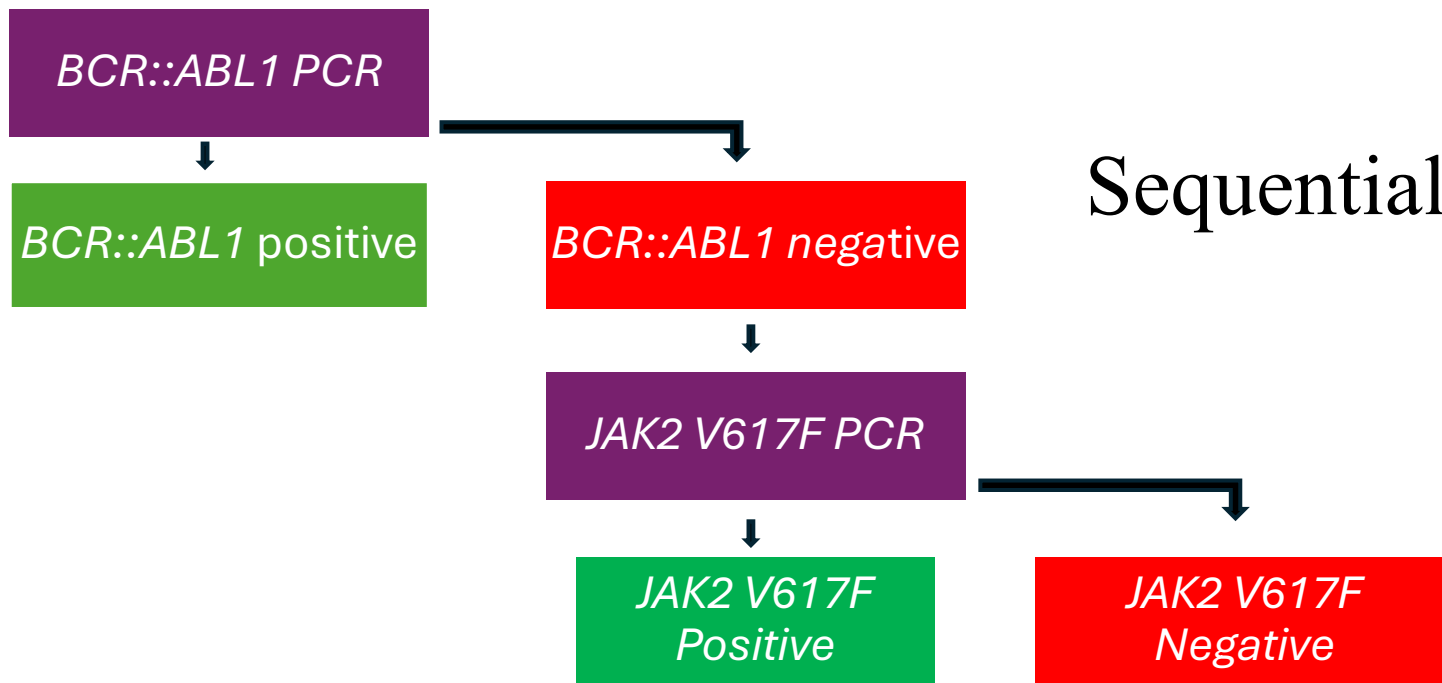


Sequential PCR based testing strategy

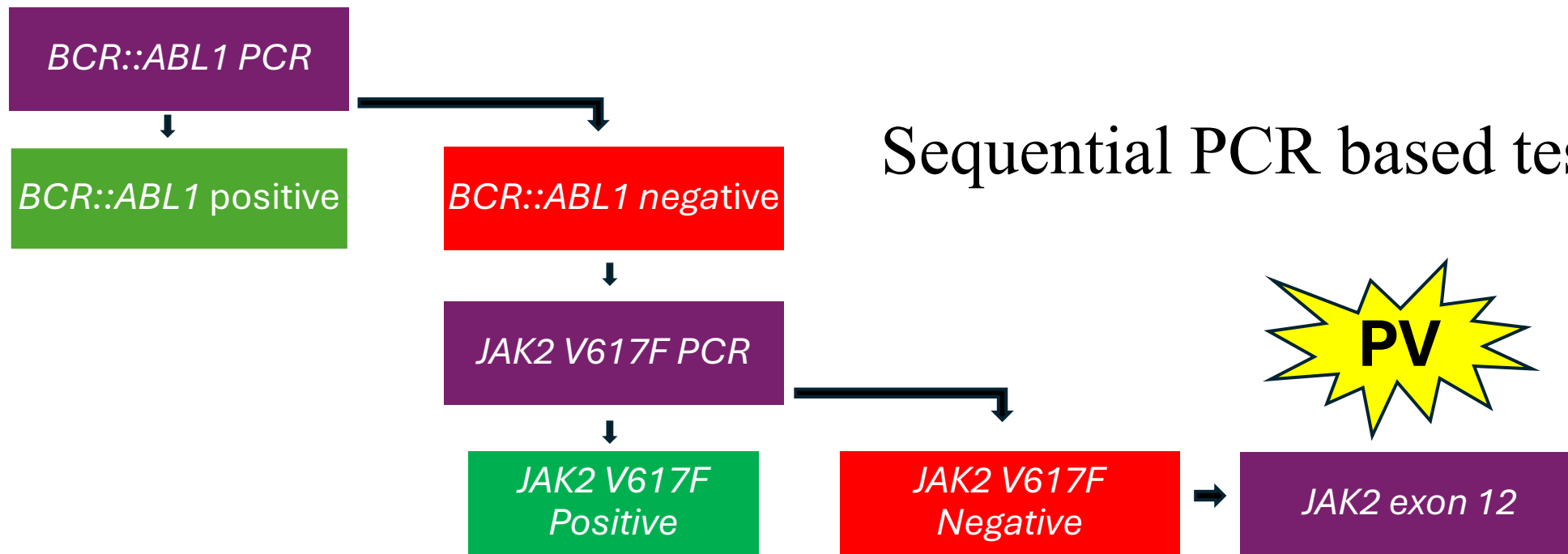
Sequential PCR based testing strategy



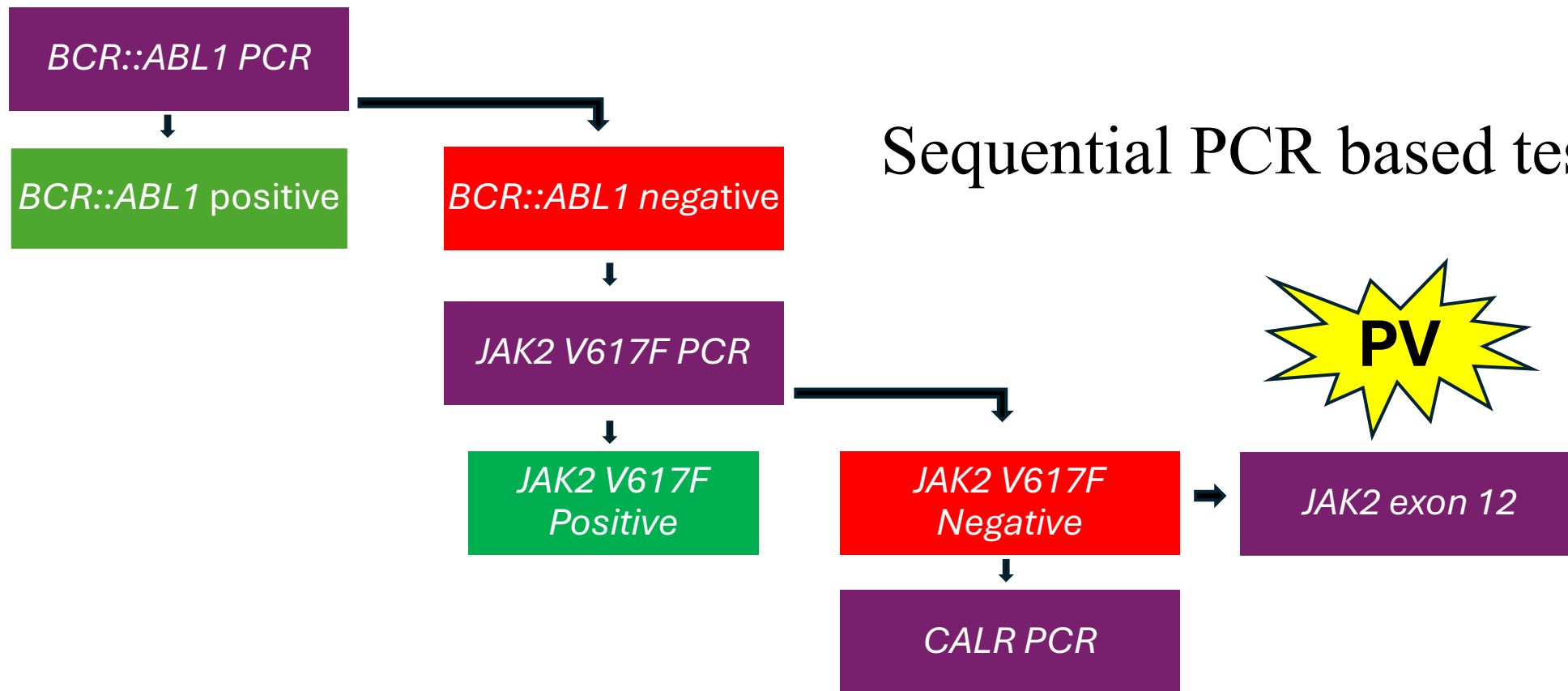
Sequential PCR based testing strategy



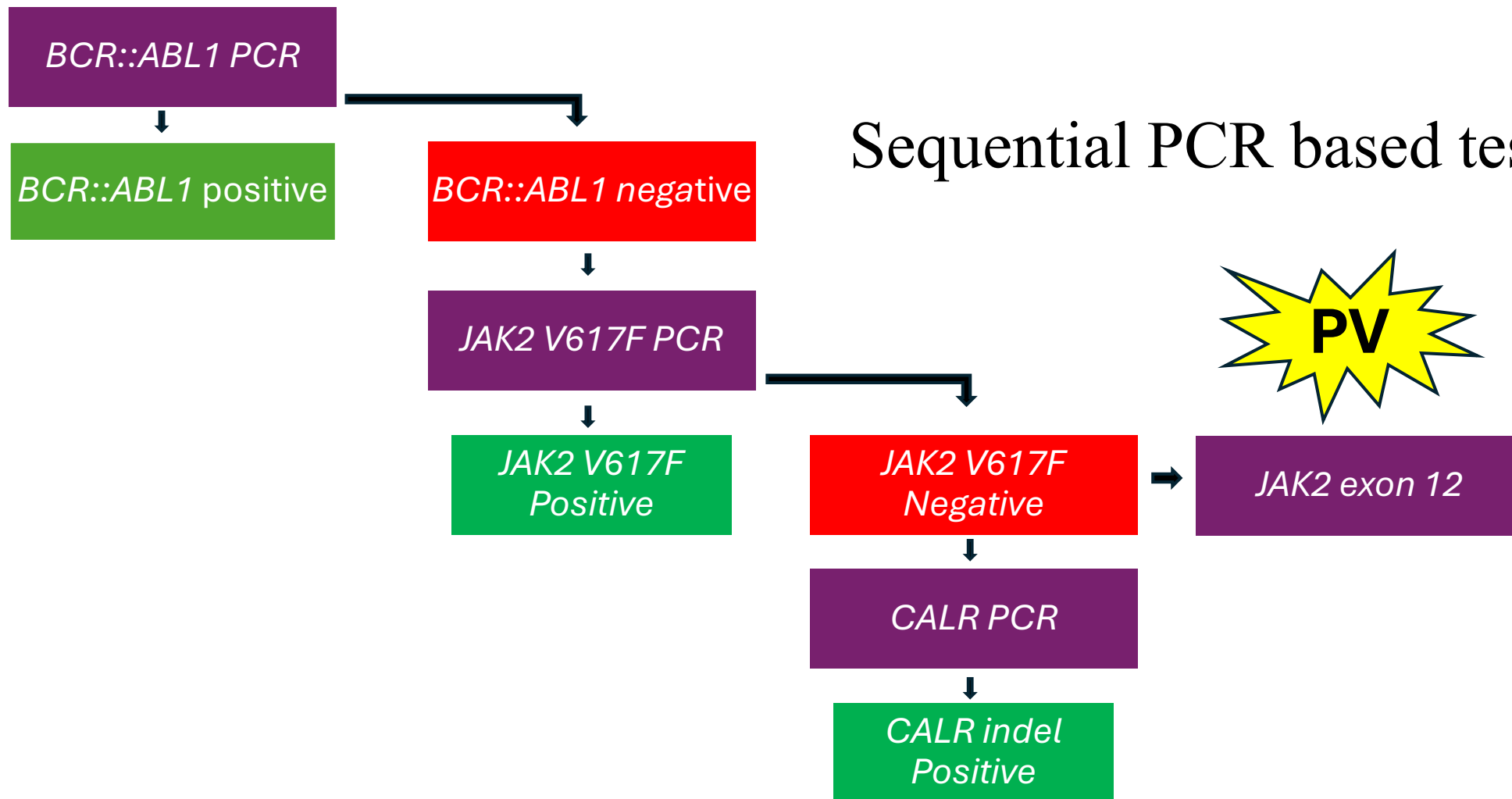
Sequential PCR based testing strategy



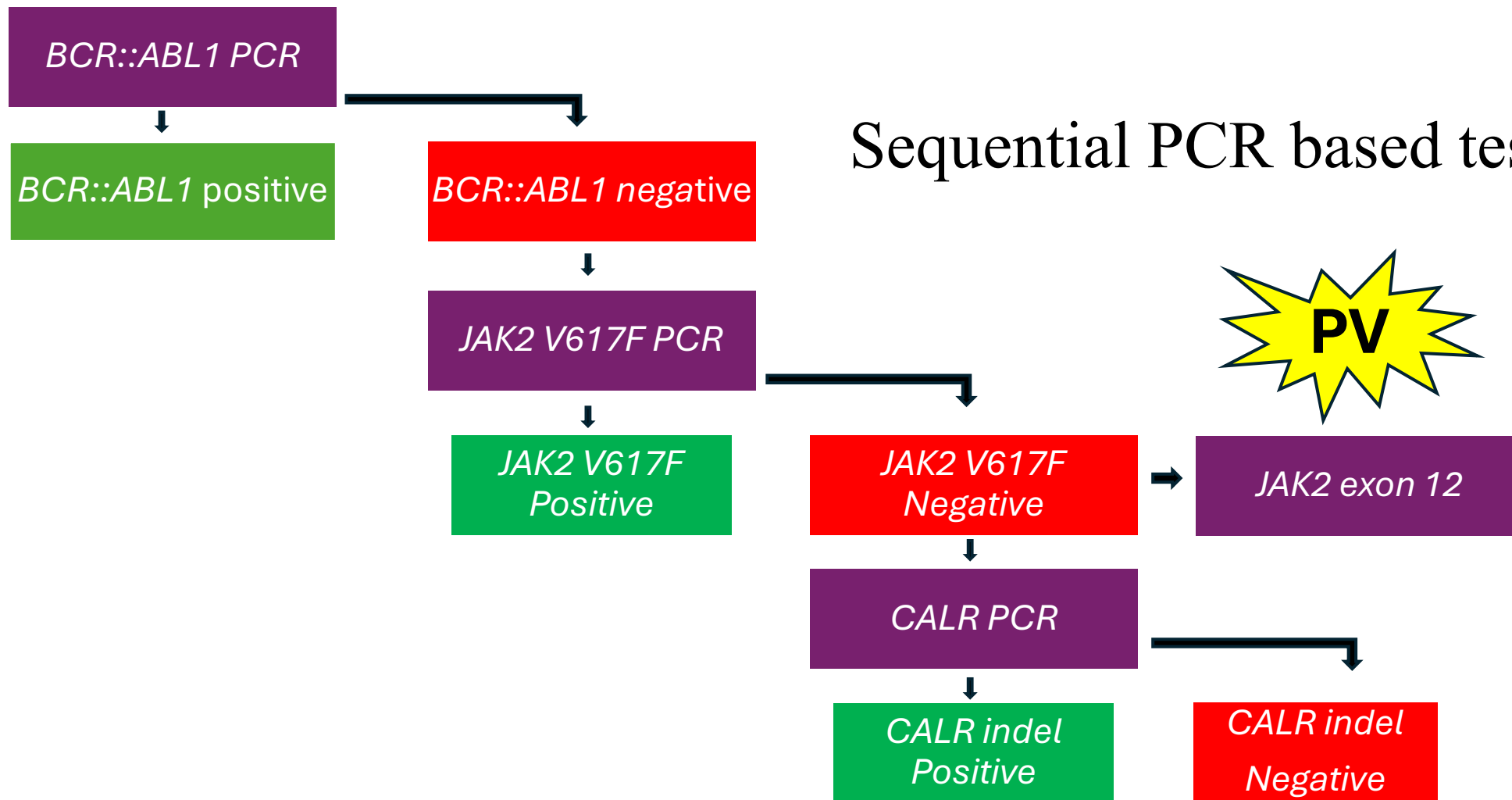
Sequential PCR based testing strategy



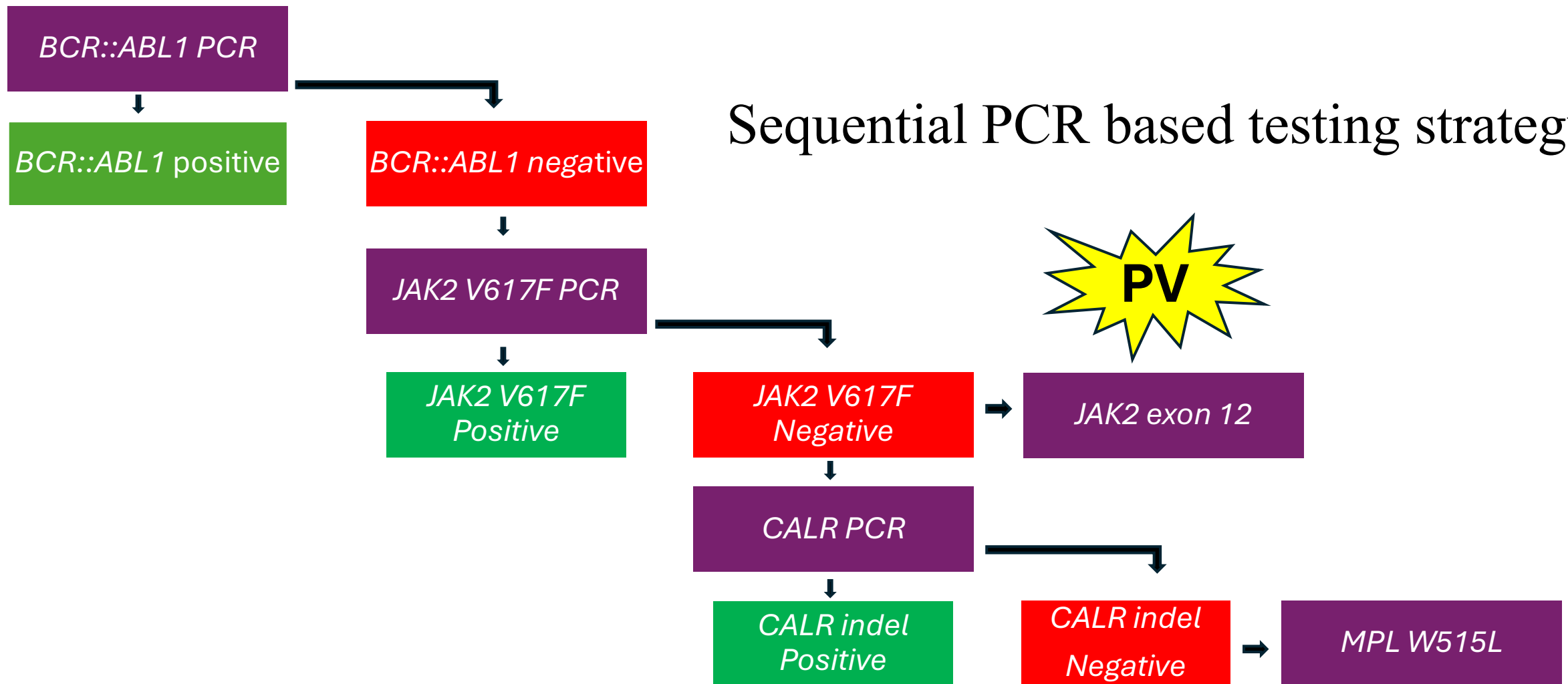
Sequential PCR based testing strategy



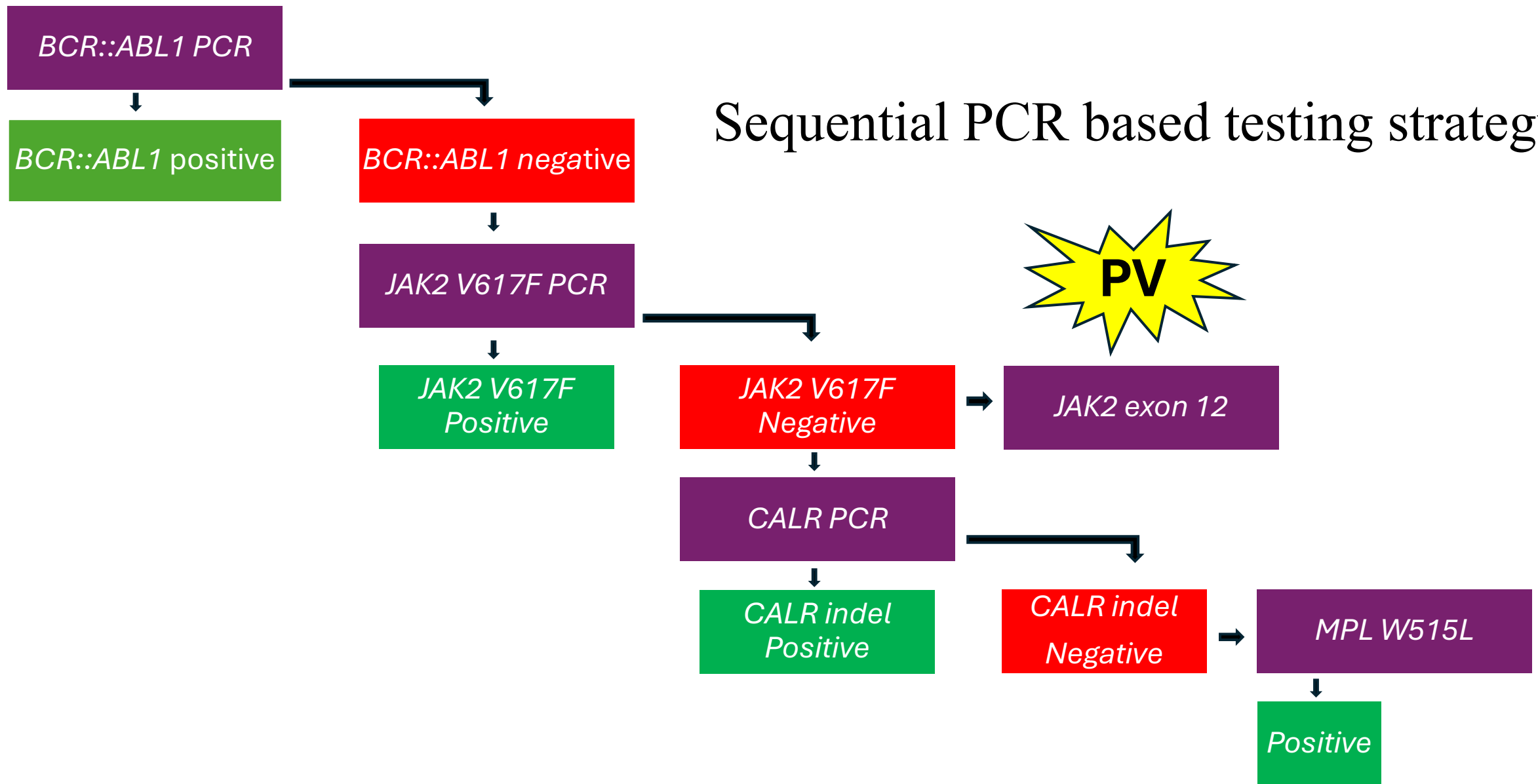
Sequential PCR based testing strategy



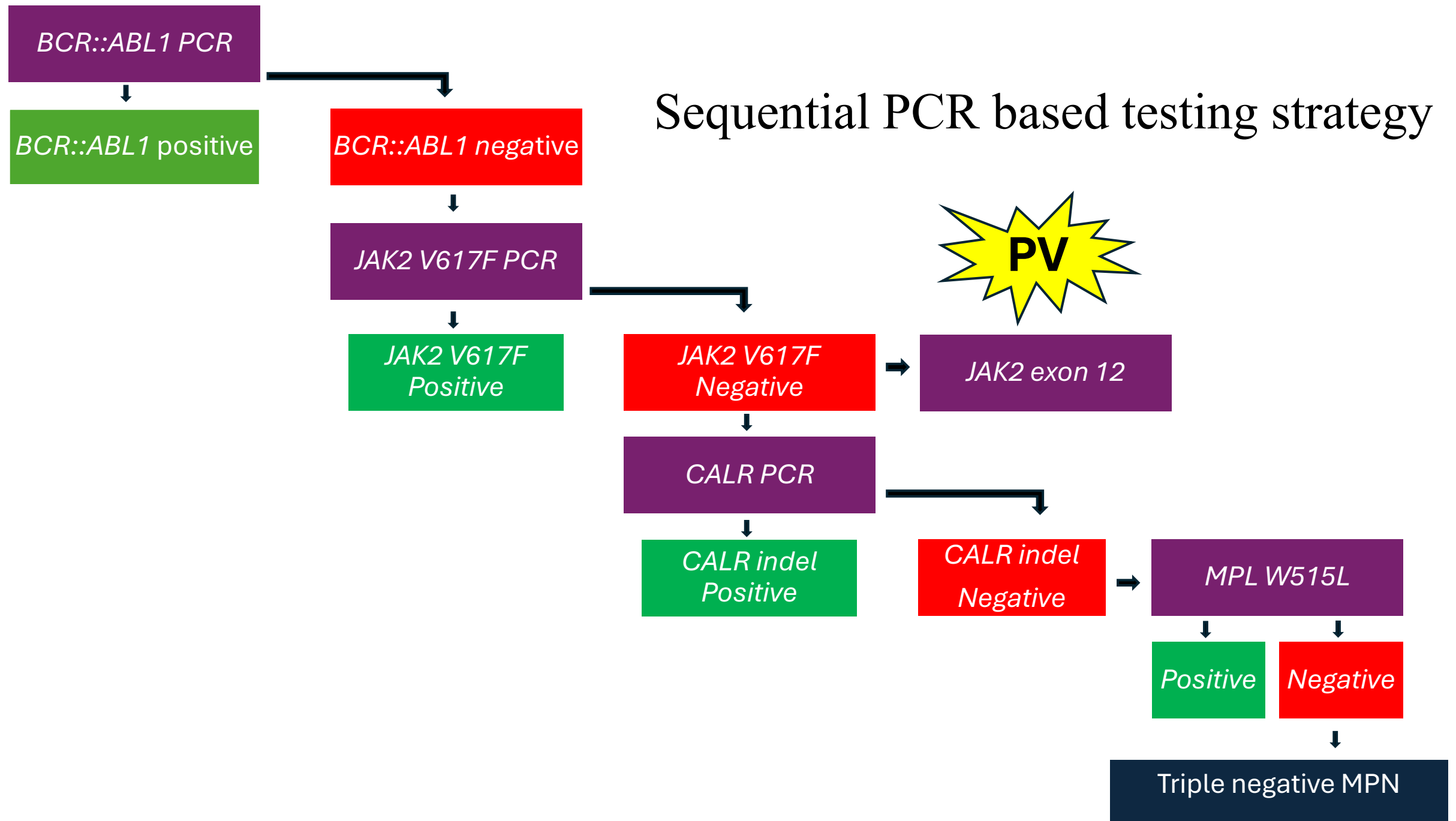
Sequential PCR based testing strategy

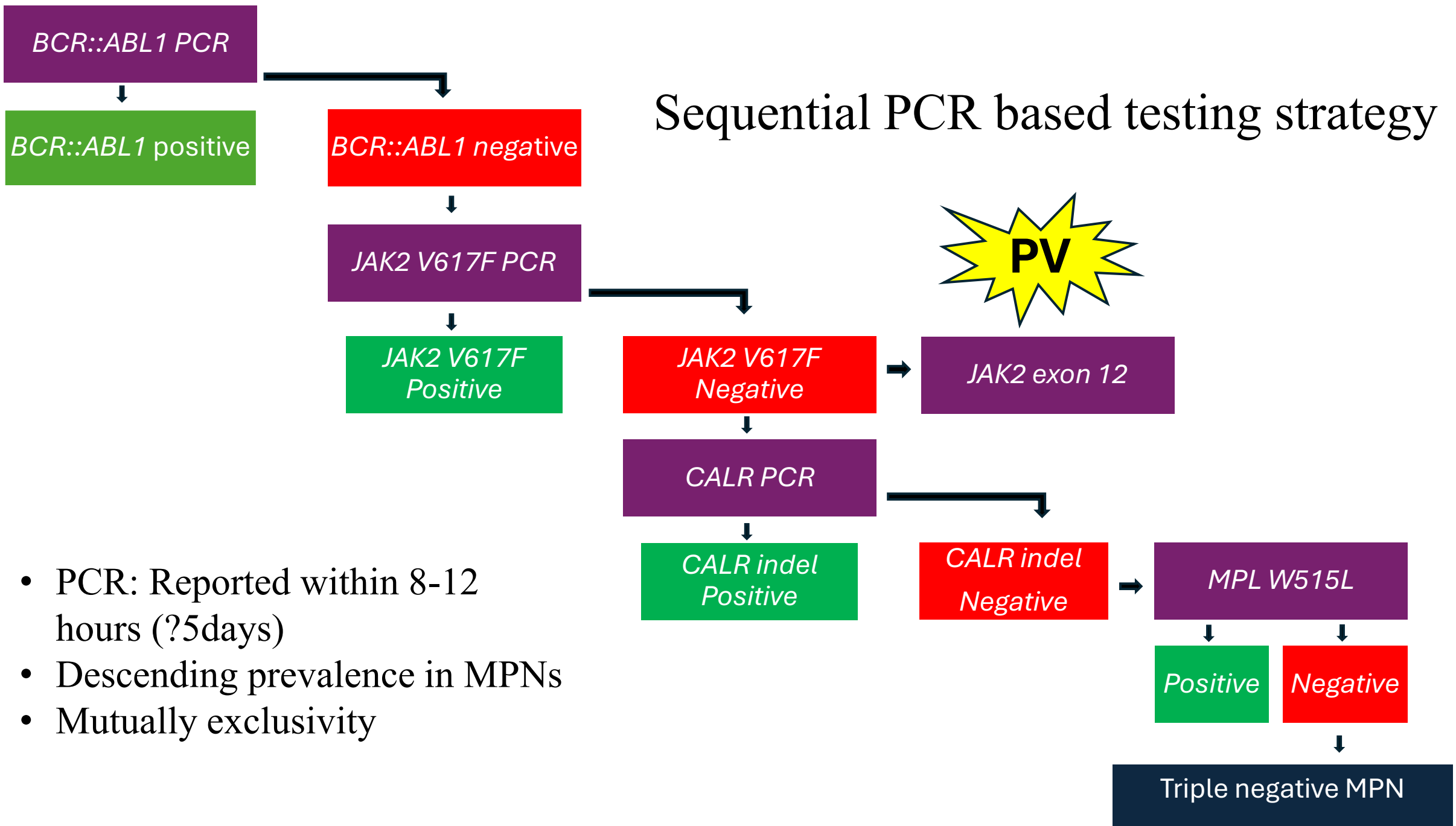


Sequential PCR based testing strategy



Sequential PCR based testing strategy





> [Sci Rep.](#) 2025 Jul 18;15(1):26046. doi: 10.1038/s41598-025-11096-6.

Clinical, morphological and genetic characteristics of patients with concurrent presence of JAK2 V617F and BCR::ABL1

Nicole Naumann¹, Vito Dangelo¹, Johannes Lübke¹, Jakob Bresser¹, Volker Hagen², Jolanta Dengler³, Georgia Metzgeroth¹, Sebastian Kreil¹, Tabea Hockenberger¹, Wolf-Karsten Hofmann¹, Alice Fabarius¹, Susanne Saussele¹, Nicholas C P Cross^{4 5}, Andreas Reiter¹, Juliana Schwaab^{6 7}

Affiliations
 + expand

PMID: 40681596

 PMCID: [PMC12274285](#)

 DOI: [10.1038/s41598-025-11096-6](#)

[Case Reports](#)
 > [J Korean Med Sci.](#) 2020 Jun 15;35(23):e168. doi: 10.3346/jkms.2020.35.e168.

A Rare Case of Essential Thrombocythemia with Coexisting *JAK2* and *MPL* Driver Mutations

Mi Ae Jang¹, Mi Yeon Seo², Kyoung Jin Choi², Dae Sik Hong³

Affiliations
 + expand

PMID: 32537949

 PMCID: [PMC7295601](#)

 DOI: [10.3346/jkms.2020.35.e168](#)

Abstract

Philadelphia-negative (Ph-) classical myeloproliferative neoplasms (MPNs) include polycythemia vera, essential thrombocythemia (ET), and primary myelofibrosis. Somatic driver mutations in the *JAK2*, *CALR*, and *MPL* genes serve as major diagnostic criteria of the Ph- MPNs and these mutations occur in a mutually exclusive manner. In this report, we describe the first case of ET harboring double mutations in *JAK2* V617F and *MPL*. For *MPL*, the patient had multiple clones of *MPL* mutations: c.1543_1546delinsAGGG (p.Trp515_Gln516delinsArgGlu) and c.1546C>G (p.Gln516Glu). The *JAK2* V617F allele burden in our patient is very low (4%) compared to the relatively high (17%-78%) allele frequency of *MPL* mutations. The low *JAK2* mutant burden might be explained by preexisting clonal hematopoiesis before overt signs of MPNs, followed by the acquisition of a second oncogenic mutation of *CALR* or *MPL* leading to the MPN phenotype. This highlights that [screening for a second driver mutation should be considered in patients with a low *JAK2* mutant burden](#) by reporting a 57-year-old Korean man with ET.

[Case Reports](#)
 > [Ann Hematol.](#) 2015 May;94(5):865-7. doi: 10.1007/s00277-014-2248-0.

Epub 2014 Nov 5.

A report on the co-occurrence of JAK2V617F and CALR mutations in myeloproliferative neoplasm patients

Na Xu¹, Li Ding, Changxin Yin, Xuan Zhou, Lin Li, Yulin Li, Qisi Lu, Xiao-li Liu

Affiliations
 + expand

PMID: 25366168

 DOI: [10.1007/s00277-014-2248-0](#)

> [Oncotarget.](#) 2016 Aug 30;7(35):57036-57049. doi: 10.18632/oncotarget.10958.

Coexistence of JAK2 and CALR mutations and their clinical implications in patients with essential thrombocythemia

Min-Gu Kang^{1 2}, Hyun-Woo Choi¹, Jun Hyung Lee¹, Yong Jun Choi¹, Hyun-Jung Choi¹, Jong-Hee Shin¹, Soon-Pal Suh¹, Michael Szardenings³, Hye-Ran Kim⁴, Myung-Geun Shin^{1 2 5}

Affiliations
 + expand

PMID: 27486987

 PMCID: [PMC5302971](#)

 DOI: [10.18632/oncotarget.10958](#)

Abstract

Janus kinase 2 (JAK2) and calreticulin (CALR) constitute the two most frequent mutations in essential thrombocythemia (ET), and both are reported to be mutually exclusive. Hence, we examined a cohort of 123 myeloproliferative neoplasm (MPN) patients without BCR-ABL1 rearrangement and additional ET patients (n=96) for coexistence of JAK2 and CALR mutations. The frequency of CALR mutations was 20.3% in 123 MPN patients; 31.1% in ET (n=74), 25% in primary myelofibrosis (n=4) and 2.2% in polycythemia vera (n=45). [JAK2 and CALR mutations coexisted in 7 \(4.2%\) of 167 ET patients](#). Clinical characteristics, progression-free survival (PFS), and elapsed time to achieve partial remission across 4 groups (JAK2+/CALR+, JAK2+/CALR-, JAK2-/CALR+, JAK2-/CALR-) were reviewed. The JAK2+/CALR- group had higher leukocyte counts and hemoglobin levels and more frequent thrombotic events than JAK2-/CALR- group. JAK2 mutations have a greater effect on the disease phenotype and the clinical features of MPN patients rather than do CALR mutation. JAK2+ groups showed a tendency of poor PFS than JAK2- groups regardless of CALR mutation. CALR+ was a predictor of late response to the treatment. Our study also showed that thrombosis was more frequent in ET patients with type 2 CALR mutations than in those with type 1 CALR mutations.

> [Am J Hematol.](#) 2018 Aug;93(4):E84-E86. doi: 10.1002/ajh.25014. Epub 2018 Jan 12.

Clinical and biological characterization of MPN patients harboring two driver mutations, a French intergroup of myeloproliferative neoplasms (FIM) study

Olivier Mansier^{1 2 3}, Damien Luque Paz^{4 5 6 7}, Jean-Christophe Ianotto^{7 8}, Yannick Le Bris^{7 9}, Aurélie Chauveau^{7 10 11 12}, Françoise Boyer^{7 13}, Carole Conejero¹⁴, Olivier Fitoussi¹⁵, Jérémie Riou¹⁶, Didier Adiko¹⁷, Mohamed Touati¹⁸, Jasmine Chauzeix^{19 20}, Jean-François Viallard²¹, Marie C Béné^{7 9}, Stéphane Giraudier¹⁴, Valérie Ugo^{4 5 6 7}, Eric Lippert^{7 10 11 12}

[Case Reports](#)
 > [Br J Haematol.](#) 2014 Oct;167(2):276-8. doi: 10.1111/bjh.12969. Epub 2014 Jun 17.

JAK2 V617F and CALR mutations are not mutually exclusive; findings from retrospective analysis of a small patient cohort

Gillian McGaffin¹, Kirsteen Harper, David Stirling, Lorna McLintock

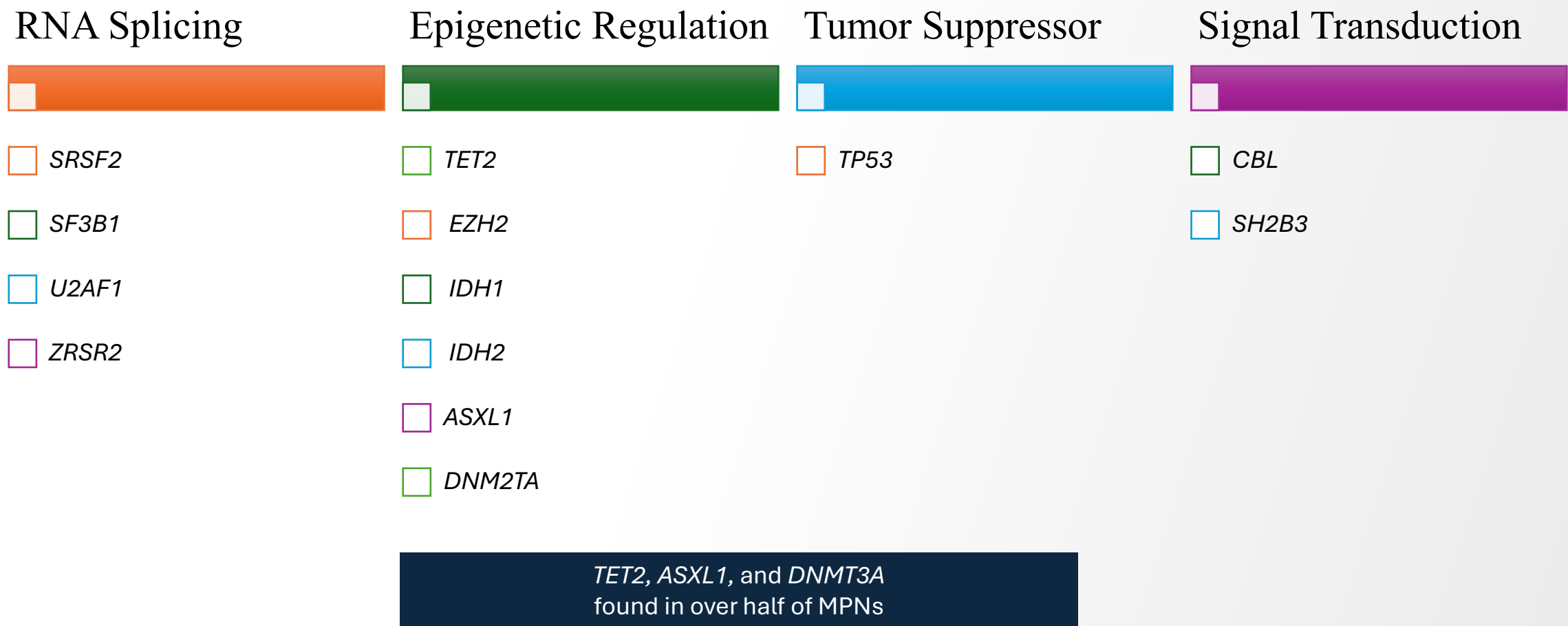
Affiliations
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PMID: 24935260

 DOI: [10.1111/bjh.12969](#)

Mutual exclusivity- Fact or Fiction?

Other MPN- Related Mutations Recommended by NCCN Guidelines



NCCN: NGS- based panel recommended to assess prognosis and establishing clonality in TN-MPNs

Primary myelofibrosis (PMF)

- *ASXL1*, *EZH2*, *SRSF2*, *TP53*, *IDH1*, *IDH2*, or *U2AF1* mutations are considered as high-molecular-risk (HMR) mutations and are associated with significantly shorter OS and leukemia-free survival (LFS) in patients with PMF.

Polycythemia Vera (PV)

- In one report, a mutation in one of these genes (*ASXL1*, *SRSF2*, and *IDH2*) was associated with inferior OS and MF-free survival but it did not significantly affect the LFS in patients with PV

Essential Thrombocythemia (ET)

- *SH2B3*, *IDH2*, *U2AF1*, *SF3B1*, *SRSF2*, *EZH2*, and *TP53* mutations were identified as significant risk factors for inferior OS, MF-free survival, and LFS in patients with ET

NGS may also be useful to establish the clonality (e.g., triple-negative MPN with non-mutated *JAK2*, *MPL*, and *CALR*)

Beyond The Three Driver Genes (ASH 2023 Publication)

- ☐ Recurrent somatic gene mutations in MPNs and secondary AML.
- ☐ Secondary AML is rare but devastating complications with a median survival of <6 months

Gene Name Location & Type of Mutation	PV %	ET %	PMF %	Post-MPN AML %	Clinical Implication
<i>TET2</i> (all exons)	10%-20%	3%-10%	10%-20%	19%-25%	No prognostic impact reported
<i>DNMT3A</i> (R882 and all exons)	5%-10%	1%-5%	8%-12%	3%-17%	No prognostic impact reported
<i>IDH1</i> (R312)	1%-2%	1%-2%	5%-6%	13%	High molecular risk (HMR) in PMF and adverse prognosis impact in all MPN subtype
<i>IDH2</i> (R140 or R172)	1%-2%	1%-2%	5%-6%	7%-15%	HMR in PMF and adverse prognosis impact in all MPN subtype
<i>ASXL1</i> (mostly nonsense/frameshift in the last exon)	2%-7%	5%-10%	15%-35%	17%-47%	HMR in PMF and adverse prognostic impact in all MPN subtypes
<i>EZH2</i> (all exons)	1%-2%	1%-2%	7%-10%	7%-13%	HMR in PMF and adverse prognosis impact in all MPN subtype
<i>NRAS</i> (G12, G13, or Q61)	<2%	<2%	2%-4%	11%	Adverse prognostic impact in all MPN subtypes
<i>KRAS</i> (G12, G13, or Q61)	<2%	<2%	2%	4%-7%	Adverse prognostic impact in all MPN subtypes
<i>SH2B3</i> (exon 2)	2%-9%	1%-3%	2%-4%	6%-11%	Rare mutations in JAK2 negative MPNs

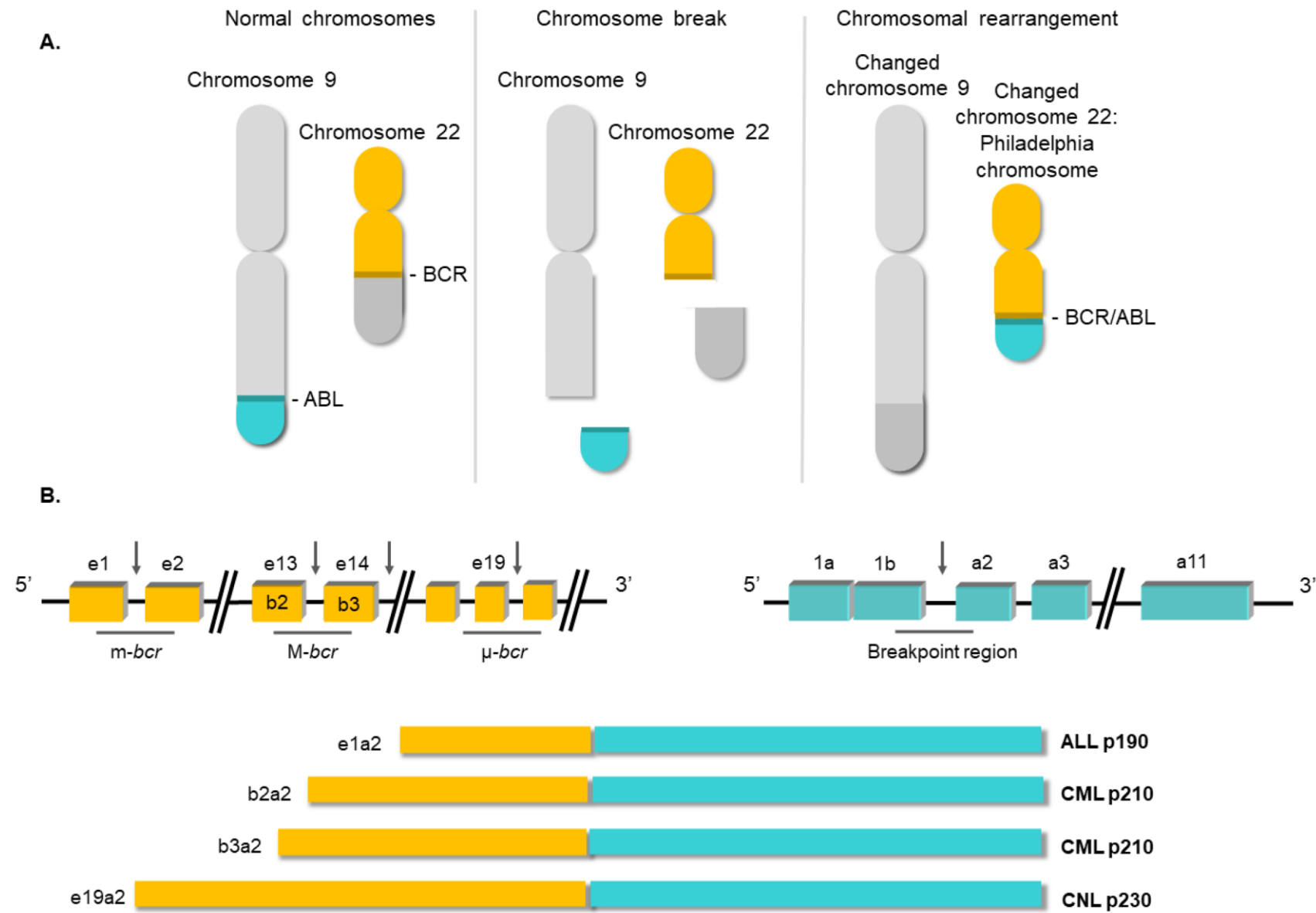
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Gene Name Location & Type of Mutation	PV %	ET %	PMF %	Post-MPN AML %	Clinical Implication
<i>CBL</i> (exon 8 and 9)	<2%	<2%	4%	4%	Adverse prognostic impact in all MPN subtypes
<i>SRSF2</i> (P95)	<2%	<2%	6%-14%	7%-22%	HMR in PMF and adverse prognostic impact in all MPN subtypes
<i>SF3B1</i> (exon 14-16)	2%-3%	2%-5%	5%-7%	7%-11%	Adverse prognostic impact in ET
<i>U2AF1</i> (S34 or q157)	<2%	<2%	7%-10%	5%-12%	Adverse prognostic impact in all MPN subtypes
<i>NFE2</i> (all exons)	3%-6%	1%-7%	3%-5%	?	Increased risk of leukemic transformation
<i>RUNX1</i> (all exons)	<2%	<2%	2%-3%	20%	Adverse prognostic impact in all MPN subtypes
<i>TP53</i> (all exons)	<2%	<2%	4%-5%	16%-50%	Adverse prognostic impact in all MPN subtypes

Luque Paz, et al. Blood 2023; 141 (16): 1909–1921. doi: <https://doi.org/10.1182/blood.2022017578>

Chronic Myeloid Leukemia (CML):



Resistance Mutations:

BCR::ABL1 kinase
domain mutations

T315I/A, M244V, G250E,
Y253H, E255K/VV299L,
F317L/V/I/C, A337T,
F359V/I/C, P465S.

BCR::ABL1 kinase
independent mutations

*ASXL1, IKZF1, BCOR,
TET1/2, IDH1/2,
DNMT3A/3B, EZH2*

AMPLICON-BASED MYELOID NGS PANEL

DNA panel: Hotspot genes (28)		DNA panel: Full genes (17)		RNA panel: Fusion driver genes (35)			RNA panel: Expression genes (5)	RNA panel: Expression control genes (5)
<i>ANKRD26</i>	<i>KRAS</i>	<i>ASXL1</i>	<i>PRPF8</i>	<i>ABL1</i>	<i>HMGA2</i>	<i>NUP98</i>	<i>BAALC</i>	<i>EIF2B1</i>
<i>ABL1</i>	<i>MPL</i>	<i>BCOR</i>	<i>RB1</i>	<i>ABL2</i>	<i>JAK2</i>	<i>NUP214</i>	<i>MECOM</i>	<i>FBXW2</i>
<i>BRAF</i>	<i>MYD88</i>	<i>CALR</i>	<i>RUNX1</i>	<i>BCL2</i>	<i>KAT6A (MOZ)</i>	<i>PAX5</i>	<i>MYC</i>	<i>PSMB2</i>
<i>CBL</i>	<i>NPM1</i>	<i>CEBPA</i>	<i>SH2B3</i>	<i>BRAF</i>	<i>KAT6B</i>	<i>PDGFRA</i>	<i>SMC1A</i>	<i>PUM1</i>
<i>CSF3R</i>	<i>NRAS</i>	<i>ETV6</i>	<i>STAG2</i>	<i>CCND1</i>	<i>KMT2A</i>	<i>PDGFRB</i>	<i>WT1</i>	<i>TRIM27</i>
<i>DDX41</i>	<i>PPM1D</i>	<i>EZH2</i>	<i>TET2</i>	<i>CREBBP</i>	<i>KMT2A PTDs</i>	<i>RARA</i>		
<i>DNMT3A</i>	<i>PTPN11</i>	<i>IKZF1</i>	<i>TP53</i>	<i>EGFR</i>	<i>MECOM</i>	<i>RUNX1</i>		
<i>FLT3 (ITD, TKD)</i>	<i>SMC1A</i>	<i>NF1</i>	<i>ZRSR2</i>	<i>ETV6</i>	<i>MET</i>	<i>TCF3</i>		
<i>GATA2</i>	<i>SETBP1</i>			<i>FGFR1</i>	<i>MLLT10</i>	<i>TFE3</i>		
<i>HRAS</i>	<i>SF3B1</i>			<i>FGFR2</i>	<i>MRTFA (MKL1)</i>	<i>ZNF384</i>		
<i>IDH1</i>	<i>SRSF2</i>			<i>FUS</i>	<i>MYBL1</i>			
<i>IDH2</i>	<i>U2AF1</i>				<i>MYH11</i>			
<i>JAK2</i>	<i>WT1</i>				<i>NTRK2</i>			
<i>KIT</i>					<i>NTRK3</i>			

AMPLICON-BASED MYELOID NGS PANEL

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<i>ABL1</i>	<i>MPL</i>	<i>BCOR</i>	<i>RB1</i>	<i>ABL2</i>	<i>JAK2</i>	<i>NUP214</i>	<i>MECOM</i>	<i>FBXW2</i>
<i>BRAF</i>	<i>MYD88</i>	<i>CALR</i>	<i>RUNX1</i>	<i>BCL2</i>	<i>KAT6A (MOZ)</i>	<i>PAX5</i>	<i>MYC</i>	<i>PSMB2</i>
<i>CBL</i>	<i>NPM1</i>	<i>CEBPA</i>	<i>SH2B3</i>	<i>BRAF</i>	<i>KAT6B</i>	<i>PDGFRA</i>	<i>SMC1A</i>	<i>PUM1</i>
<i>CSF3R</i>	<i>NRAS</i>	<i>ETV6</i>	<i>STAG2</i>	<i>CCND1</i>	<i>KMT2A</i>	<i>PDGFRB</i>	<i>WT1</i>	<i>TRIM27</i>
<i>DDX41</i>	<i>PPM1D</i>	<i>EZH2</i>	<i>TET2</i>	<i>CREBBP</i>	<i>KMT2A PTDs</i>	<i>RARA</i>		
<i>DNMT3A</i>	<i>PTPN11</i>	<i>IKZF1</i>	<i>TP53</i>	<i>EGFR</i>	<i>MECOM</i>	<i>RUNX1</i>		
<i>FLT3 (ITD, TKD)</i>	<i>SMC1A</i>	<i>NF1</i>	<i>ZRSR2</i>	<i>ETV6</i>	<i>MET</i>	<i>TCF3</i>		
<i>GATA2</i>	<i>SMC3</i>	<i>PHF6</i>		<i>FGFR1</i>	<i>MLLT10</i>	<i>TFE3</i>		
<i>GATA2</i>	<i>SETBP1</i>			<i>FGFR2</i>	<i>MRTFA (MKL1)</i>	<i>ZNF384</i>		
<i>HRAS</i>	<i>SF3B1</i>			<i>FUS</i>	<i>MYBL1</i>			
<i>IDH1</i>	<i>SRSF2</i>				<i>MYH11</i>			
<i>IDH2</i>	<i>U2AF1</i>				<i>NTRK2</i>			
<i>JAK2</i>	<i>WT1</i>				<i>NTRK3</i>			
<i>KIT</i>								

= MPN-related genes

AMPLICON-BASED MYELOID NGS PANEL

Missing : *NFE2*

DNA panel: Hotspot genes (28)		DNA panel: Full genes (17)		RNA panel: Fusion driver genes (35)			RNA panel: Expression genes (5)	RNA panel: Expression control genes (5)
<i>ANKRD26</i>	<i>KRAS</i>	<i>ASXL1</i>	<i>PRPF8</i>	<i>ABL1</i>	<i>HMGA2</i>	<i>NUP98</i>	<i>BAALC</i>	<i>EIF2B1</i>
<i>ABL1</i>	<i>MPL</i>	<i>BCOR</i>	<i>RB1</i>	<i>ABL2</i>	<i>JAK2</i>	<i>NUP214</i>	<i>MECOM</i>	<i>FBXW2</i>
<i>BRAF</i>	<i>MYD88</i>	<i>CALR</i>	<i>RUNX1</i>	<i>BCL2</i>	<i>KAT6A (MOZ)</i>	<i>PAX5</i>	<i>MYC</i>	<i>PSMB2</i>
<i>CBL</i>	<i>NPM1</i>	<i>CEBPA</i>	<i>SH2B3</i>	<i>BRAF</i>	<i>KAT6B</i>	<i>PDGFRA</i>	<i>SMC1A</i>	<i>PUM1</i>
<i>CSF3R</i>	<i>NRAS</i>	<i>ETV6</i>	<i>STAG2</i>	<i>CCND1</i>	<i>KMT2A</i>	<i>PDGFRB</i>	<i>WT1</i>	<i>TRIM27</i>
<i>DDX41</i>	<i>PPM1D</i>	<i>EZH2</i>	<i>TET2</i>	<i>CREBBP</i>	<i>KMT2A PTDs</i>	<i>RARA</i>		
<i>DNMT3A</i>	<i>PTPN11</i>	<i>IKZF1</i>	<i>TP53</i>	<i>EGFR</i>	<i>MECOM</i>	<i>RUNX1</i>		
<i>FLT3 (ITD, TKD)</i>	<i>SMC1A</i>	<i>NF1</i>	<i>ZRSR2</i>	<i>ETV6</i>	<i>MET</i>	<i>TCF3</i>		
<i>GATA2</i>	<i>SMC3</i>	<i>PHF6</i>		<i>FGFR1</i>	<i>MLLT10</i>	<i>TFE3</i>		
<i>GATA2</i>	<i>SETBP1</i>			<i>FGFR2</i>	<i>MRTFA (MKL1)</i>	<i>ZNF384</i>		
<i>HRAS</i>	<i>SF3B1</i>			<i>FUS</i>	<i>MYBL1</i>			
<i>IDH1</i>	<i>SRSF2</i>				<i>MYH11</i>			
<i>IDH2</i>	<i>U2AF1</i>				<i>NTRK2</i>			
<i>JAK2</i>	<i>WT1</i>				<i>NTRK3</i>			
<i>KIT</i>								

= MPN-related genes

Case	Clinical presentation	Notable laboratory data	JAK2 V617F PCR	CALR exon 9 PCR	JAK2 exon 12 test	MPL test	NGS result (blood)
Case 1	64-y-old man with persistent thrombocytosis for 2 y	RBC count: $4.15 \times 10^{12}/L$ Hemoglobin: 135 g/L Hematocrit: 42.3% WBC count: $5.4 \times 10^9/L$ Platelet count: $489 \times 10^9/L$	+	-	N/A	N/A	JAK2 V617F (VAF = 0.9%)*; MPL W515L (VAF = 2.3%)
Case 2	65-y-old man with leukocytosis and thrombocytosis; hypersegmented neutrophils and giant platelets noted	RBC count: $3.76 \times 10^{12}/L$ Hemoglobin: 111 g/L Hematocrit: 33.9% WBC count: $27.1 \times 10^9/L$ Platelet count: $979 \times 10^9/L$	+	-	N/A	N/A	JAK2 V617F (VAF = 46.1%); IDH2 R140W (VAF = 2.2%)
Case 3	85-y-old man with thrombocytosis	RBC count: $3.43 \times 10^{12}/L$ Hemoglobin: 105 g/L Hematocrit: 31.0% WBC count: $14.1 \times 10^9/L$ Platelet count: $2553 \times 10^9/L$	+	-	N/A	N/A	JAK2 V617F (VAF = 30.7%); SF3B1 K666T (VAF = 35.5%)
Case 4	57-y-old woman with primary erythrocytosis	RBC count: $5.32 \times 10^{12}/L$ Hemoglobin: 162 g/L Hematocrit: 49.7% WBC count: $6.4 \times 10^9/L$ Platelet count: $248 \times 10^9/L$	-	-	-	N/A	DNMT3A R882H (VAF = 5.7%)
Case 5	63-y-old man with borderline erythrocytosis and thrombocytosis	RBC count: $5.67 \times 10^{12}/L$ Hemoglobin: 166 g/L Hematocrit: 48.7% WBC count: $8.2 \times 10^9/L$ Platelet count: $466 \times 10^9/L$	-	-	-	N/A	DNMT3A Q527* (VAF = 2.5%); TET2 Q1852* (VAF = 1.9%)
Case 6	55-y-old woman with erythrocytosis	RBC count: $5.47 \times 10^{12}/L$ Hemoglobin: 162 g/L Hematocrit: 49.5% WBC count: $7.3 \times 10^9/L$ Platelet count: $308 \times 10^9/L$	-	-	-	-	DNMT3A N838D (VAF = 2.8%)

Case #1:

64 y/o male with persistent thrombocytosis for two years

Notable Laboratory Data

RBC: $4.15 \times 10^{12}/L$
HB: 135 g/L
HCT: 42.3%
WBC: 5.4×10^9
PLTs: $489 \times 10^9/L$ 

PCR Results

JAK2 V617F (+)
CALR (-)
JAK2 exon 12 (NA)
MPL (NA)

Amplicon – based myeloid NGS panel (PB)

JAK2 V617F (VAF =0.9%)

MPL W515L (VAF=2.3%)

Case #1:

64 y/o male with persistent thrombocytosis for two years

Notable Laboratory Data

RBC: $4.15 \times 10^{12}/L$
HB: 135 g/L
HCT: 42.3%
WBC: 5.4×10^9
PLTs: $489 \times 10^9/L$ 

PCR Results

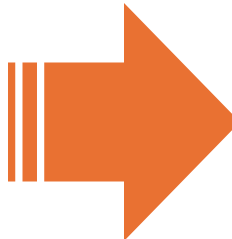
JAK2 V617F (+)
CALR (-)
JAK2 exon 12 (NA)
MPL (NA)

Amplicon – based myeloid NGS panel (PB)

JAK2 V617F (VAF =0.9%)

MPL W515L (VAF=2.3%)

Screening for a second driver mutation in low *JAK2*V617F allele burden (<1%): The disease phenotype of dictated by the predominant mutation



Case #2:

65 y/o male with leukocytosis & thrombocytosis; hypersegmented neutrophils and giant platelets

Notable Laboratory Data

RBC: $3.76 \times 10^{12}/L$
HB: 111 g/L
HCT: 33.9%
WBC: $27.1 \times 10^9/L$
PLTs: $979 \times 10^9/L$



PCR Results

JAK2 V617F (+)
CALR (-)
JAK2 exon 12 (NA)
MPL (NA)

Amplicon- based myeloid NGS panel (PB)

JAK2 V617F (VAF =46.1)

IDH2 R140W (VAF=2.2%)

Case #2:

65 y/o male with leukocytosis & thrombocytosis; hypersegmented neutrophils and giant platelets

Notable Laboratory Data

RBC: $3.76 \times 10^{12}/L$
HB: 111 g/L
HCT: 33.9%
WBC: $27.1 \times 10^9/L$
PLTs: $979 \times 10^9/L$

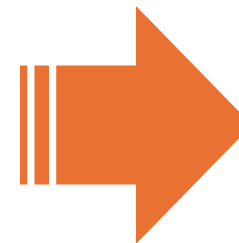
PCR Results

JAK2 V617F (+)
CALR (-)
JAK2 exon 12 (NA)
MPL (NA)

Amplicon- based myeloid NGS panel (PB)

JAK2 V617F (VAF =46.1)

IDH2 R140W (VAF=2.2%)



High molecular risk and
adverse prognostic impact in
MPNs.
Treatment with IDH inhibitors?

Case #3:

85 y/o male presented with thrombocytosis

Notable Laboratory Data

RBC: 3.43×10^{12}
HB: 105 g/L
HCT: 31.0%
WBC: 14.1×10^9
PLTs: 2553×10^9



PCR Results

JAK2 V617F (+)
CALR (-)
JAK2 exon 12 (NA)
MPL (NA)

Amplicon- based myeloid NGS panel (PB)

JAK2 V617F (VAF =30.7)

SF3B1 K666T (VAF=2.2%)

Case #3:

85 y/o male presented with thrombocytosis

Notable Laboratory Data

RBC: 3.43×10^{12}
HB: 105 g/L
HCT: 31.0%
WBC: 14.1×10^9
PLTs: 2553×10^9



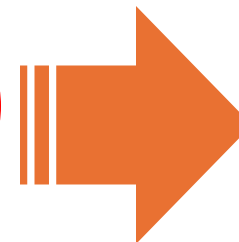
PCR Results

JAK2 V617F (+)
CALR (-)
JAK2 exon 12 (NA)
MPL (NA)

Amplicon-based myeloid NGS panel (PB)

JAK2 V617F (VAF = 30.7)

SF3B1 K666T (VAF = 2.2%)



A gain-of-function mutation associated with inferior overall and myelofibrosis-free survival in patients with ET

Case #4:

57 y/o female with primary erythrocytosis

Notable Laboratory Data

RBC: $5.32 \times 10^{12}/L$
HB: 162 g/L
HCT: 49.7%
WBC: $6.4 \times 10^9/L$
PLTs: $248 \times 10^9/L$



PCR Results

JAK2 V617F (-)
CALR (-)
JAK2 exon 12 (-)
MPL (NA)

Amplicon- based myeloid NGS panel (PB)

DNMT3A R882H (VAF=5.7%)

Case #4:

57 y/o female with primary erythrocytosis

Notable Laboratory Data

RBC: $5.32 \times 10^{12}/L$
HB: 162 g/L
HCT: 49.7%
WBC: $6.4 \times 10^9/L$
PLTs: $248 \times 10^9/L$

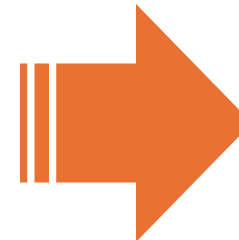


PCR Results

JAK2 V617F (-)
CALR (-)
JAK2 exon 12 (-)
MPL (NA)

Amplicon- based myeloid NGS panel (PB)

DNMT3A R882H (VAF=5.7%)



DNMT3A mutations are
well reported in CHIP
and myeloid neoplasms

Case #5:

63 y/o male with borderline erythrocytosis and thrombocytosis

Notable Laboratory Data

RBC: $5.67 \times 10^{12}/L$
HB: 166 g/L
HCT: 48.7%
WBC: $8.2 \times 10^9/L$
PLTs: $466 \times 10^9/L$



PCR Results

JAK2 V617F (-)
CALR (-)
JAK2 exon 12 (-)
MPL (NA)

Amplicon- based myeloid NGS panel (PB)

DNMT3A Q527 (VAF=2.5%)

TET2 Q1852 (VAF = 1.9%)

Case #5:

63 y/o male with borderline erythrocytosis and thrombocytosis

Notable Laboratory Data

RBC: $5.67 \times 10^{12}/L$
HB: 166 g/L
HCT: 48.7%
WBC: $8.2 \times 10^9/L$
PLTs: $466 \times 10^9/L$

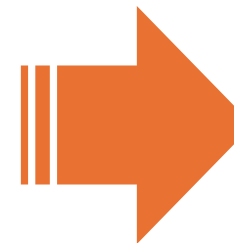
PCR Results

JAK2 V617F (-)
CALR (-)
JAK2 exon 12 (-)
MPL (NA)

Amplicon- based myeloid NGS panel (PB)

DNMT3A Q527 (VAF=2.5%)

TET2 Q1852 (VAF = 1.9%)



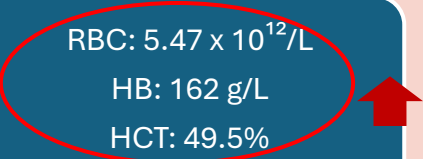
DNMT3A and *TET2* variants
could provide additional
information in terms of
thrombotic risk prediction
and disease prognosis

Case #6:

55 y/o female with erythrocytosis

Notable Laboratory Data

RBC: $5.47 \times 10^{12}/L$
HB: 162 g/L
HCT: 49.5%
WBC: $7.3 \times 10^9/L$
PLTs: $308 \times 10^9/L$



PCR Results

JAK2 V617F (-)
CALR (-)
JAK2 exon 12 (-)
MPL (-)

Amplicon- based myeloid NGS panel (PB)

DNMT3A N838D (VAF=2.8%)

Case #6:

55 y/o female with erythrocytosis

Notable Laboratory Data

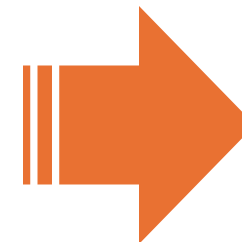
RBC: $5.47 \times 10^{12}/L$
HB: 162 g/L
HCT: 49.5%
WBC: $7.3 \times 10^9/L$
PLTs: $308 \times 10^9/L$

PCR Results

JAK2 V617F (-)
CALR (-)
JAK2 exon 12 (-)
MPL (-)

Amplicon- based myeloid NGS panel (PB)

DNMT3A N838D (VAF=2.8%)

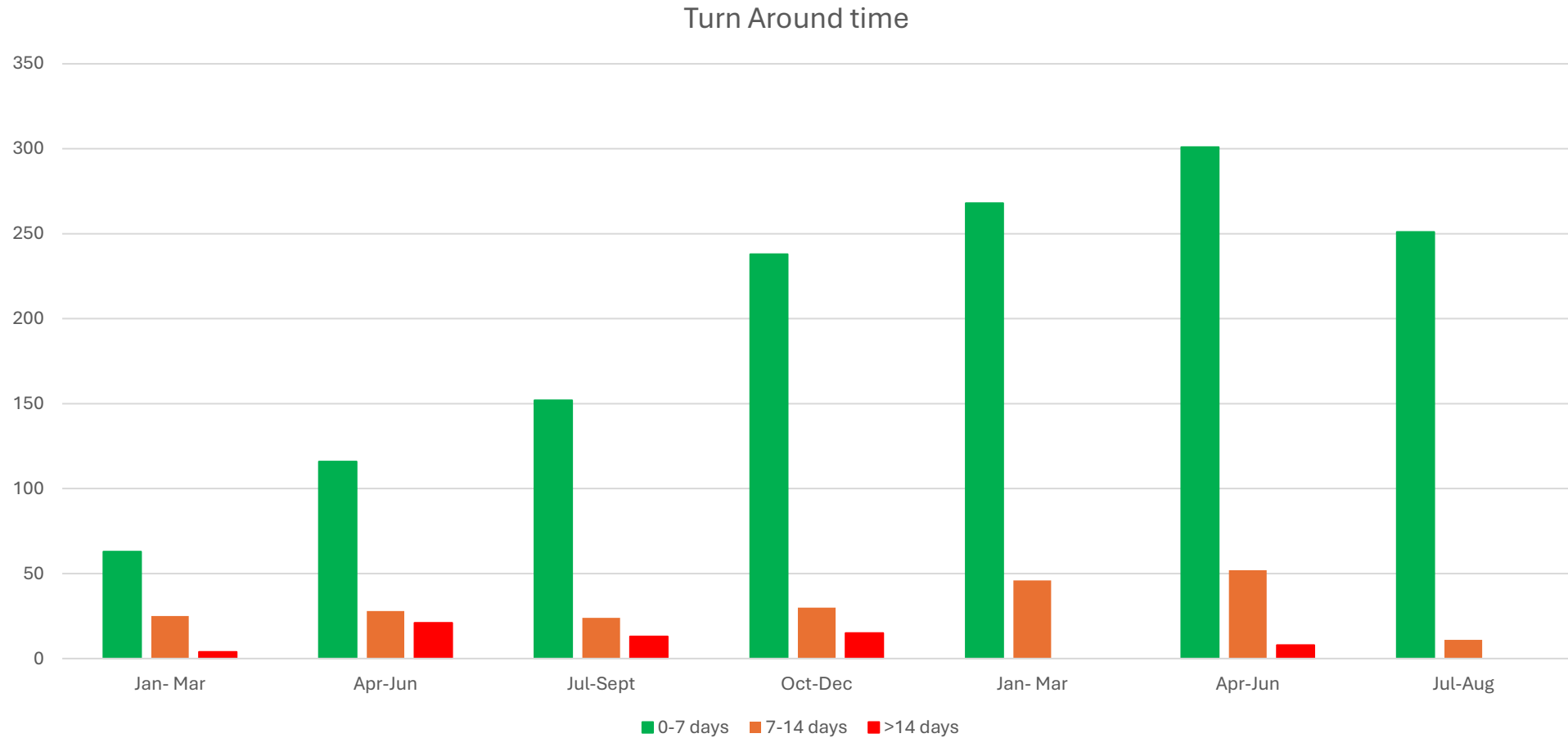


DNMT3A mutations are
well reported in CHIP
and myeloid neoplasms

NGS is a Win Win?????



Turn around times for NGS

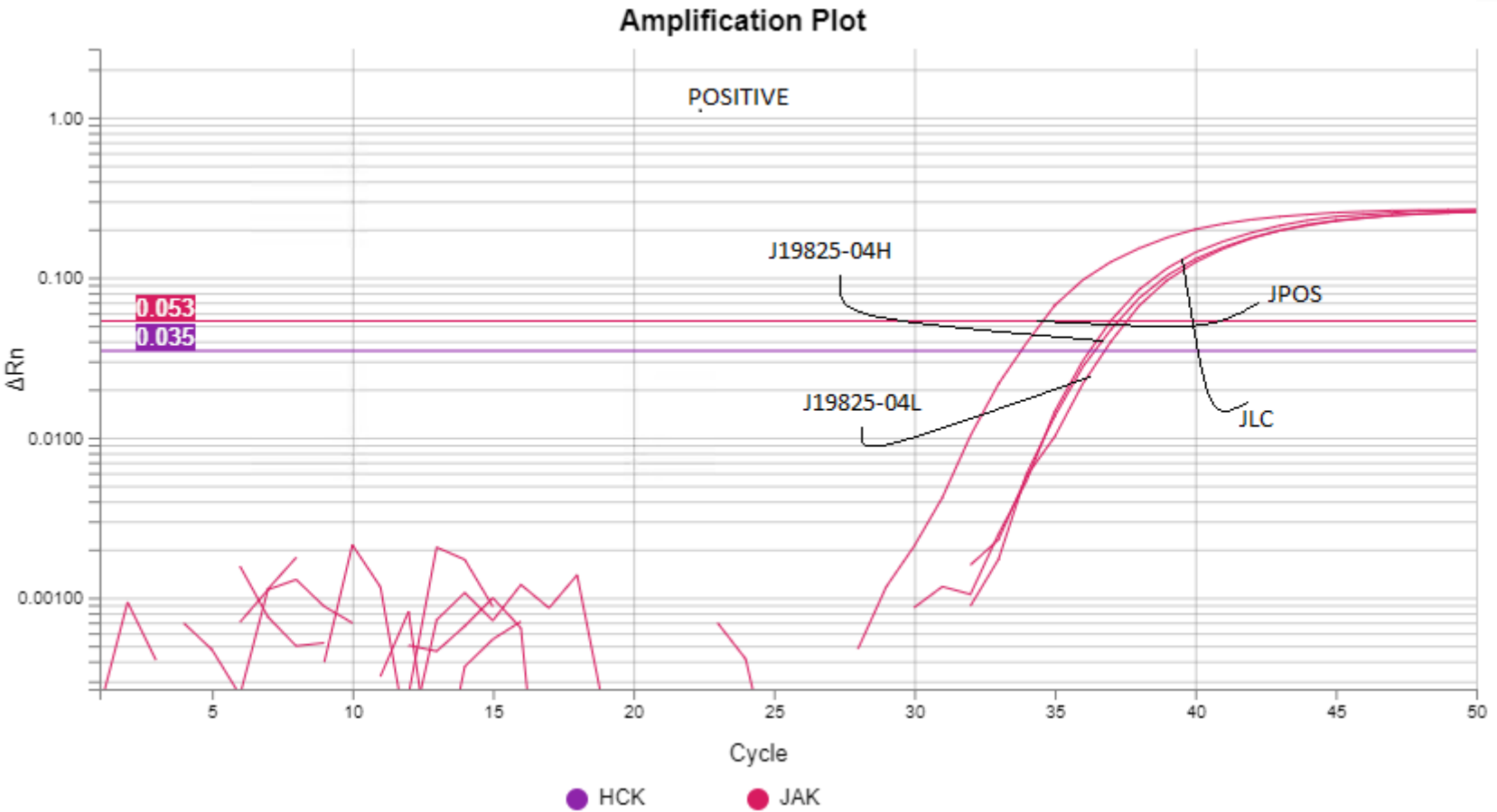




All	SNVs/Indels	Fusions							Filter Chain Applied 2 of 2,599 Variants	Edit Filters	Export ▾	Columns ▾
User Classification	Variant ID	Variant Name	Key Variant			Locus						
Classification ▾	.	DNMT3A C557W	No	No		chr2:25467204						
Classification ▾	KMT2A-KMT2A.K9K2	KMT2A PTD	Yes	No		chr11:118355029 - chr11:118339490	RNAExonVariant					Gain-of-Funct

All	SNVs/Indels	Fusions					Filter Chain Applied 2 of 2,599 Variants	Edit Filters	Export	Columns
User Classification	Variant ID	Variant Name	Key Variant		Locus					
Classification	.	DNMT3A C557W	No	No	chr2:25467204					
Classification	KMT2A-KMT2A.K9K2	KMT2A PTD	Yes	No	chr11:118355029 - chr11:118339490	RNAExonVariant	Gain-of-Funct			

Discrepant JAK2
V617F PCR?



AllSNVs/IndelsFusions

No Filter Chain Applied
1,741 of 1,741 SNVs/Indels

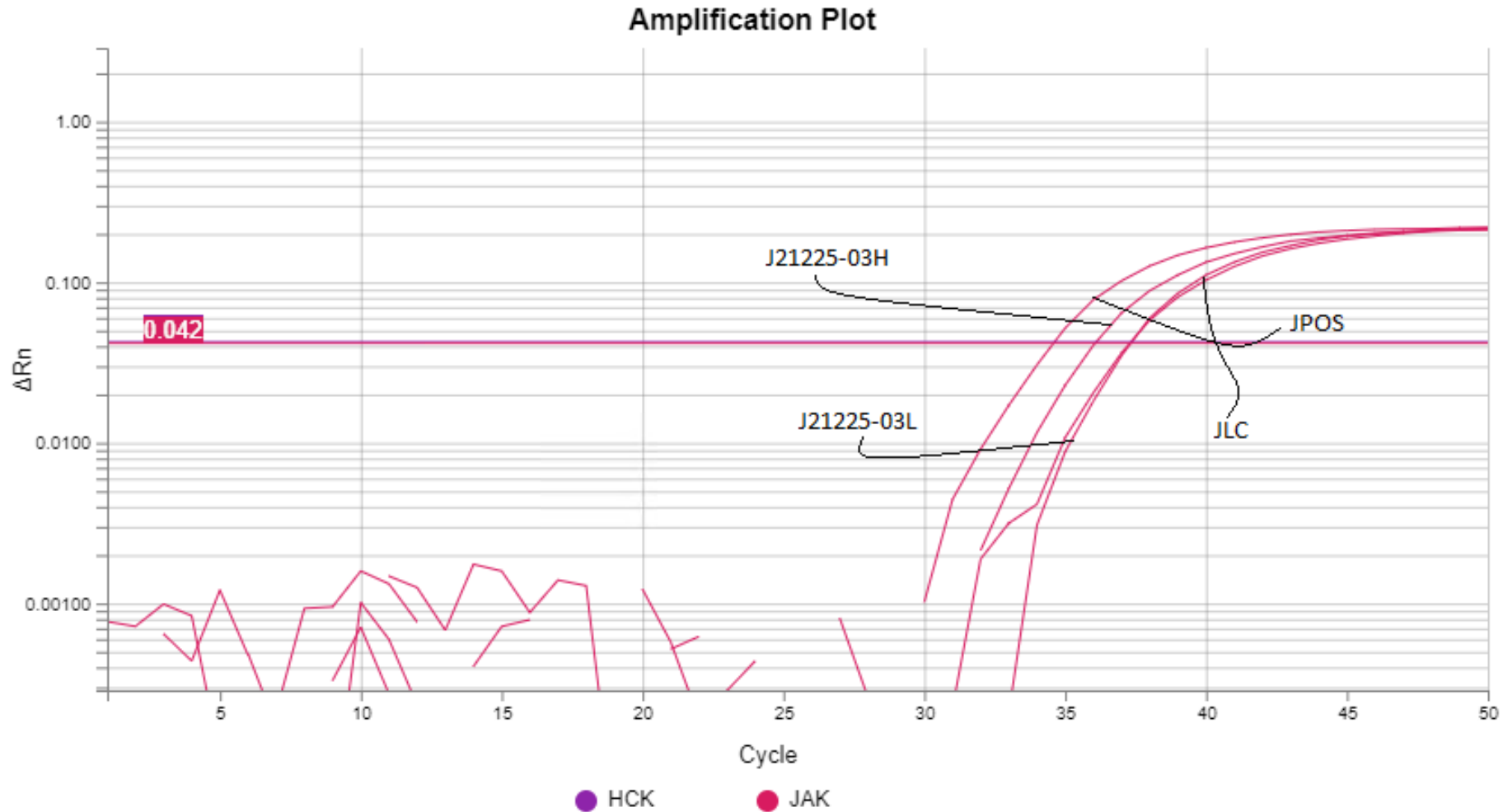
Edit Filters

Variant ID	Variant Name	Locus	Gene	AA Change	Call	Effective Read Depth	Nuc Change	Allele Frequency (%)	Allele read Count
COSM12600	JAK2 V617F	chr9:5073770	JAK2	p.Val617Phe	NO CALL	1987	c.1849G>T	1.31	

** More than one gene is annotated for this variant. Click on the variant row to view the annotation details.

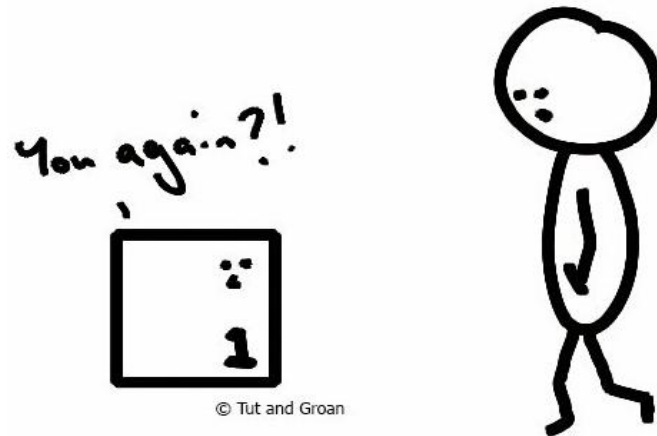


Follow up PCR 6 weeks later

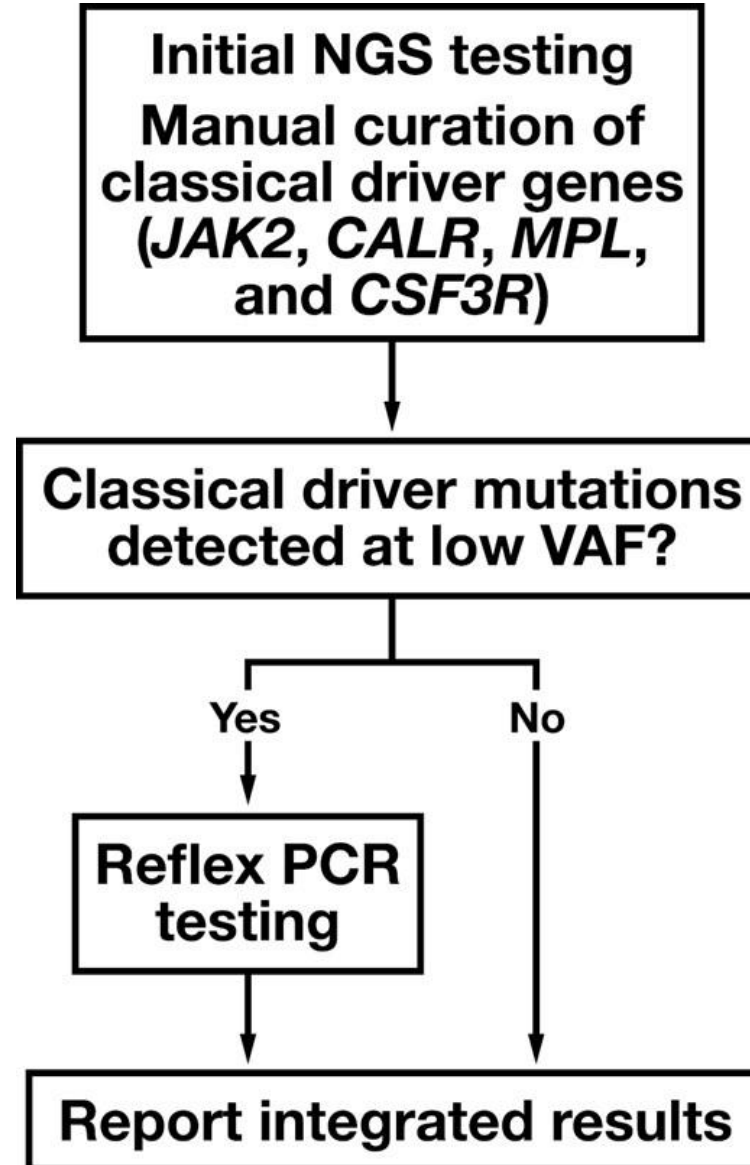


- Sensitivity of NGS for detecting low allele-frequency variants is limited by the proportional representation of these variants within the amplified product
- PCR-based testing and targeted amplification ensures that even low frequency variants are preferentially amplified.

Back to Square One



INCORPORATING
THE ADVANTAGES
OF NGS-BASED
AND PCR-BASED
TESTING



MPN Workup

Diagnosis

Proposed integrated algorithm

Prognosis

Proposed integrated algorithm

Therapy Selection

Proposed integrated algorithm

**Disease Monitoring
(MRD)**

Proposed integrated algorithm



THANK YOU

- Dr Goldstein
- Dr Naeem
- Dr KH Ramesh
- Dr Xiaowei Liu
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- All our wonderful technologists,
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