REFERENCE INTERVAL DETERMINATION IN THE LABORATORY

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Disclosures

- I have nothing to disclose in relation to this presentation
- No products will be discussed in my presentation.

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Objectives:

- Identify components in laboratory testing that will impact reference intervals
- Analyze data collection and determine the statistics to best represent your patient population that you are testing.
- Differentiate between verification and validation of a reference interval.
- Apply recommendations from guidelines for reference range determination.

Reference Intervals

- One of the most important studies conducted in the laboratory
- Up to 80% of medical decisions are made based on laboratory test results
- Problematic because many laboratories lack the time, resources, finances and in many cases the expertise to conduct these studies
- Many reference intervals are obtained from either package inserts or from publications

History

- Grasbek and Fellman published a paper in the 1950's entitled 'Normal Values and Statistics' as an initial study in the field of reference intervals (RIs)
- It was determined several years later that the terminology of 'normal values' was not adequate and even partially incorrect
- Reference values came into use from 1987 to 1991.
- The International Federation of Clinical Chemistry (IFCC) published a series of 6 papers, in which it was recommended that each laboratory follow defined procedures to produce its own reference values
- EP28 by CLSI Defining, Establishing, and Verifying Reference Intervals in the Clinical Laboratory, 3rd Edition was published 2010

Responsibility of the laboratory:

- Mandated for laboratories to publish reference intervals alongside patient test results.
- Directives from laboratory regulatory authorities have stated that no matter what size or resources that they need to justify the reference levels they adopt.
- ISO 15189:2007 the international standard that defines quality and competency in clinical laboratories state that "reference intervals shall be periodically reviewed" and verified every time a variation in analytical and/or preanalytical procedure takes place.
- In the US, Clinical Laboratory Improvements Amendments (CLIA) from 2003 states that when FDA-approved test systems are adopted unmodified, laboratories should "verify that the manufacturers' reference intervals are appropriate for the laboratory's patient population"

Current Laboratory practices: CAP Q- probe studies:

- Surveyed 500 laboratories and noted that 78% used manufacturers published ranges
- Participating laboratories were asked to supply their adult and pediatric reference intervals (low and high limits) for four common clinical chemistry parameters (potassium, calcium, magnesium and TSH) and three equally common hematological parameters (hemoglobin, platelet count and activated partial thromboplastin time).
- Asked when and how these reference intervals were arrived at, how long since they were last reviewed and the measuring platform for each analyte.
- Survey results revealed that a range of approaches were used to arrive at selected reference intervals.
- Only a half of the laboratories reported analyzing samples from healthy individuals for adult reference intervals.

Current practices

- The most frequent external source was manufacturers' recommendations/package inserts
- Even fewer (25 %) reported analyzing samples in preparation of pediatric reference intervals. The remaining laboratories adopted reference intervals from external sources without any internal study.
- Q probe studies of 163 laboratories revealed only 5.5% of labs could recall the year they revalidated their aPTT reference interval
- In another cohort of 116 laboratories, 42% established their own RI for PT and aPTT however only half did it for every change of reagent lot

Current practices:

- Among those laboratories that conducted any sort of internal study, the number of samples analyzed ranged from as few as 20 to >100.
- The results of sample analysis were used to establish reference intervals in around a half of these laboratories.
- For the remaining laboratories, results of the internal study were used to validate externally sourced reference intervals.
- 26 % of the participating laboratories do not have a written policy for establishing, revising or updating reference intervals.
- Approximately two thirds of the laboratories reported that they had revalidated their reference intervals in the year that a new analyzer was purchased
- Some laboratories reported no validation of reference intervals in the previous 10 years
- One case there had been no validation for at least 22 years.

Analysis of the data:

- Most laboratories (80%) there was "only slight" variation in reference interval limits. However, in the remaining 20 % of laboratories, a substantial variation was evident,
- Statistical analysis of the whole data set (reference intervals from all laboratories for all seven analytes) revealed that of 1271 adult reference intervals 40 (3.1 %) contained at least one limit that was a statistical outlier.
- For some of the analytes (magnesium, TSH and APTT) a certain amount of the observed variation in reference intervals between laboratories could be accounted for by differences in analytical methodology, but it certainly did not account for all of the variation.
- Inaccurate reference intervals may be being used to interpret patient test results.

What do laboratories actually do for reference intervals?

- Strict adherence for reference ranges for the most commonly requested analytes,
- Less stringent selection of the reference sample population (blood donors or patients without problems likely to affect the analyte) were used for more esoteric analytes,
- Specific literature on reference values, particularly individual publications with data obtained using the laboratories methodology,
- General literature concerning reference values, particularly compendia from professional bodies or Standard Operating Procedure or clinical guidelines.
- Manufacturers' data as quoted in technical data sheets and package inserts.

Review of Terminology:

- Observed value: value of an analyte obtained by observation or measurement of a test subject, which should be compared with reference values, a reference distribution, reference limit or reference interval
- Reference distribution: the distribution of reference values
- Reference individual: a person selected on the basis of well-defined criteria
- **Reference population**: a group consisting of all reference individuals
- Reference interval: the interval between two reference limits (these included) e.g.: 95% of apparently healthy men from 18 to 65 years
- Reference limits: a value derived from the reference distribution and used for descriptive purposes
- Reference values: the value obtained by observation or measurement of a defined quantity on a reference individual

The concept is as follows:

- Reference individuals
- Make up a reference population
- From whom are selected a reference sample group
- On whom are determined reference values
- On which is observed a reference distribution
- From which are determined reference limits
- That define a reference interval.

Reference Limits

- The *reference limits*: (defining a reference range)
 - Associated with a well-defined reference population,
 - Generally consisting of healthy individuals
 - Used to compare an observed value (a result from the patient) to reference data obtained from this group of well-defined subjects.
 - One of the keys for medical decision making which should take into account the specificities of each patient.
 - Descriptive of a given health state

Medical Decision Limits

■ The medical decision limits

Used by the clinician as a threshold below or above which a medical action is recommended.

Reference limits are generally two (upper and lower limits), the number of decision limits is variable according to concerned laboratory test and clinical setting. They are based on a clinical assessment and are set either by statistical methods (e.g. Bayesian approach) or from epidemiological studies.

Clinical Decision Limits

- For some analytes, reference ranges are replaced by decision limits set by national or international consensus (e.g. total cholesterol, glycated hemoglobin)
- They are determined using receiver operator curves (ROCs) that define the probability of disease.
- For these analytes it is unnecessary to determine reference limits or to validate data from the literature.
- Reference values are calculated specific to health whereas CDLs indicate sensitivity to disease

Selection of Subjects



What is "normal"?

- Adults
- Pediatrics
- Geriatrics
- **■** Ethnicities
- Gender

What is a reference interval?

Reference Intervals are defined in relation to a healthy population to include the values in which 95% of apparently healthy individuals would fall and in which 2.5 % of results in the lower range are out of the RI and 2.5% of values in the upper range will be out of the RI.

Selection of subjects

- A questionnaire should be used to capture relevant information including health status, age, gender, ethnicity and any medicationsincluding OTC, vitamins and supplements
- Equal amounts of male/female
- Representative of patient testing population
- A priori or a posteriori approach
- The more defined the population the better the outcomes

Biological Variation

- Defined as the variation seen in an individual subject when they are measured for an analyte repeatedly over time.
- Variations can be influenced by stress, APR, circadian rhythms and seasonal variations
- Difference is the between subject or inter- individual variation
- Studies are limited
- Most RI testing is conducted in the morning, patient blood draws occur throughout the day- clinic samples tend to arrive later in the day
- Does this impact RI reflection on patient values.

Fraser C. Biological Variation: From Principles to Practice. AACC Press 2001.

Consequences of biological variation for reference intervals: Creatinine

- Study looked at 13 were women and 13 men.
- Results showed:
 - 1. No individual had test results that spanned the entire reference interval and the results from each individual occupied only a small part of the reference interval,
 - 2. The means for most individuals lay within the reference interval and were different from each other,
 - 3. A few individuals had results which spanned the lower reference limit and these individual had values which changed from "normal" to "abnormal" (as clinicians would usually say) over time, and
 - 4. A few individuals had results that spanned the upper reference limit and these individuals also had values that changed from normal to abnormal over time.

https://acutecaretesting.org/en/articles/biological-variation-and-reference-normal-values

Studies on Biological Variation:

- Chen et al: 16 coag tests (PT, aPTT, fib, TT, FDP, II, V, VI, VII, VIII, IX and X)
- n=31- tested on days 1,3, and 5 at 8am, 12pm and 4pm.
- All screening CVI < 5% exception of fibrinogen
- CVI for coagulation factors >5%
- Lupus, AT, PC, S, Vw (n=25) CV < 3%

Chen Q, et al Semin Thromb Hemost 2015;41:336-41.

Shou W, Chen Q, et al Semin Thromb Hemost. 2016;42:87-92.

What information from Biological variations

- Can aid in the evaluation of results obtained in RI
- If a large variation in results are seen for a particular test, it may be worthwhile to understand if the results may be caused by known BV for that analyte.

Age specific ranges

- Newborn and childrens' systems are immature in comparison to adults and older children
- Basing pediatric results on adult ranges can cause a result to be misclassified as abnormal
- Generate a diagnosis or treatment that may not be warranted.

Problems with these ranges:

- Obtaining blood from neonates and children problematic
- Parental consent
- Insufficient volumes for testing
- Laboratories used published data
- Manufacturers published ranges
- Ranges should be adapted from the same instrument/reagent combinations

Ethnic ranges do we need them?

- Serum creatinine has different distributions for African-Americans and Europeans
- Estimated glomerular filtration rate (eGFR), a widely-used test of kidney function, is a calculation based on four variables: serum creatinine, age, sex, and race.
- In this test, race is binary (African-American or European), and recent work suggests that more granular categories and/or genetic ancestry could improve eGFR scaling

Pediatrics Reference Intervals Coagulation testing:

Study of 218 healthy children stratified by age:

II, IX, XI and XII significantly decreased in the youngest children (< 12 months)

PC and PS decreased in young childhood

Highest levels vWF in youngest children, but not FVIII

■ ARUP study- n=902 7-17 yrs old: each group n=164

PT testing 1 second longer than adults, aPTT not significant

Also confirmed age dependent ranges in FVIII, IX and XI and vW testing (activity and antigen)

Data mining; using EMR's

- Hard to obtain populations- geriatric and pediatric
- Use of EMR- larger amount of data- stratification for age groups
- Global reference intervals over a large regional area
- Expanded sample size
- Difficult to control pre-analytical variables.
- This approach is acceptable only if the laboratorian is able to identify healthy individuals not affected by a disease. This method is not recommended by the CLSI-IFCC.

Gollomp K, Arulselvan A, Tanzer M, Shibutani S, Lambert MP, Blood. 2015;126:4450

EMR ranges: Pediatrics (n=265)

- Patients (n=265) were excluded based on diagnosis or medications that could impact coagulation testing results
- Established PT= 11.6-13.8 sec post EMR review: PT=12.9-13.9 sec
- aPTT = 22-35 sec

 Post EMR review aPTT=25-35 sec
- No difference in gender was found
- Age: 2-11 yrs. PT=12.5-13.6 sec 12-23 yrs. PT=13.05-13.9 sec with no significant difference in aPTT.
- Important to understand the CV of the individual test- to determine if these results are significant
- Results may be statistically significant, but not clinically significant.

Number of subjects



Validation versus Verification

- Validation requires a minimum of 120 subjects
- Verification can use as little as 20 subjects to demonstrate test performance from a previous claim.
- Statistical evaluation of a RI are based on the number of subjects used.
- Performed:

Change in reagent

Lot number

Instrument

Collection system

CLSI document H47-A2; 2008.

Validation of a parameter that is gender dependent:

- To establish a reference interval for hemoglobin a test that is gender dependent the laboratory would need to obtain hemoglobin results on 240 reference individuals (120 men and 120 women).
- These individuals are typically recruited from the general regional population (essentially the facilities' market-base) and then selected for inclusion in the study using carefully defined criteria.
- The selection is often accomplished by administering a health questionnaire. Sometimes a physical examination is also required as a way to determine acceptability for inclusion.

Statistics:

- The RI is the range between an upper and lower limit which represents a percentage of the population tested.
- The mean and the standard deviation (SD) can be calculated. The SD is the spread of data around the mean.
- The more dispersed the data, the higher the deviation.

Confidence interval

- Measures the level of uncertainty
- 95% CI: ranges are measured at 2.5 and 97.5 percentiles of the distribution of results contains the true mean of 95% of the population
- 99% CI ensures 99% certainty
- The higher the confidence levels at the wider the CI
- Increasing the CI from 95-99% to ensure the interval contains the population mean will increase the sample size
- Using a smaller number of RI data should be statistically evaluated to see if they fit normal distribution or if it is skewed.
- Skewed data needs to be normalized by log transformation prior to calculating and converting it back

Statistical methods for evaluation:

- 1. Parametric method: used when population is normal or Gaussian, if not a statistical transformation to normalize the data is applied
- 2. Non-parametric method: used when careful subject selection and sufficient number (120) data is collected, doesn't require laws of probability
- 3. Robust method: used in a limited sample size without requiring a Gaussian distribution- measures the position and dispersion instead of mean and SD. Sorts the data from lowest to highest in equal parts and looks at how far values are distributed from the center

Gaussian Distribution

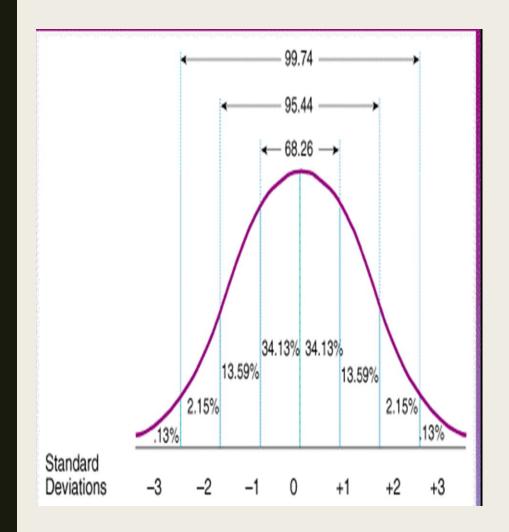
- If test results from a normal healthy patient population fall into a bell-shaped, Gaussian, normal distribution, the central 95% is usually used as the test's normal range.
- For many (but not all) tests, this is how the range of tests results for normal healthy individuals is determined.

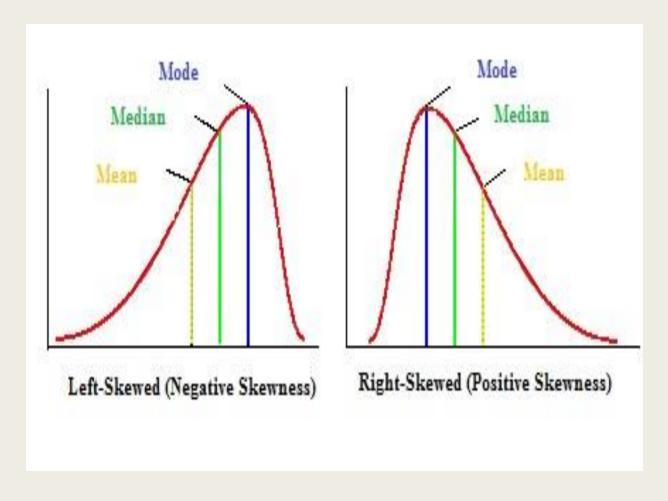
Some Normals are Abnormal and Vice Versa

- The normal range encompasses the mean plus or minus two standard deviations or, again, about 95% of normal, healthy individuals' test results.
- However, 5% (roughly 1 out of 20) **normal healthy patients** may be **outside** the cutoff value.
 - Roughly 2.5% of normal people can be expected to have a result below and roughly 2.5% of normal people can be expected to have a result above the reported normal range.
- This situation is encountered with almost all tests.
- This is because the distribution of tests results from normal, healthy individuals overlaps with the distribution of test results from sick patients with the relevant disease.

Non-Gaussian Distributions

- For non-Gaussian distributions, lab directors can use other nonparametric techniques to establish reference range limits
 - Example: set upper and lower limits of normal to include 95% of the population after all of the test results have been transformed into logarithms taking the central 95% of the transformed data.



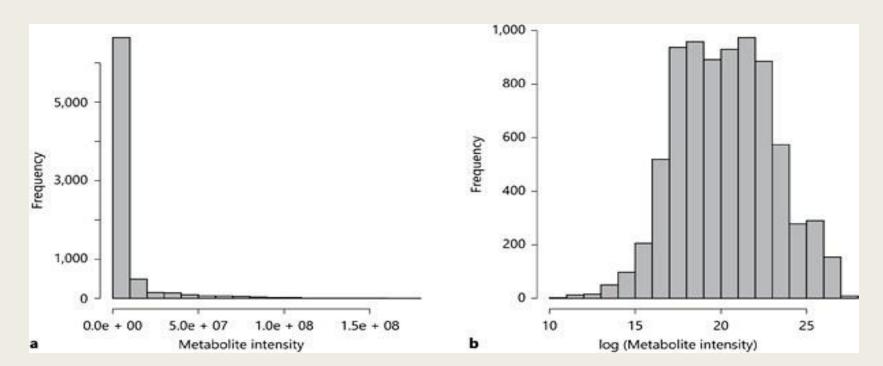


Gaussian distribution

Skewed Data

Transformation of data

- Replacement of a variable by either the square root or the logarithm of the variable changes the shape of a distribution or relationship
- Done when data is skewed to transform the data into symmetrical distribution



Outliers

- Method by Dixon: Looks at the difference in the point which appears to be an outlier and the next observation- (D)
- This is divided by difference of the lowest observation and the highest observation (R)
- If the D/R ration is greaten than 1/3, this point may be an outliers.
- In a sample size of 20, allowed 2 outliers
- If > 2 outliers, must test an additional 20 normal
- If another 2 outliers other sources of errors (reagents, analyzers, biological variations) should be investigated
- A full RI may have to be conducted.

Recommendations & Guidelines:



Transference

- Validation of a RI conducted by a manufacturer
- Smaller number of subjects n=20
- Compared to a larger study
- Keeping in mind the importance in using the same analytical test system and reagents
- If a laboratory changes methods and the method comparison is compatible, the RI can be transferred

Transference

- A laboratory can elect to "transfer" the RIs that were in use with an older method (or from another laboratory) to a new method.
- To do this, the laboratory must first demonstrate that the 2 methods produce comparable results.
- It is well known that analytic systems drift over time, and there is no guarantee that the method of today is producing results that are comparable to those that were produced at the time of the original RI study.
- This technique is the main reason why many laboratories today are using RIs that were established decades ago and are out-of-date.

Transferring existing reference intervals

- Normal laboratory practice is to perform a method comparison study in which the same fresh patient samples are measured by both methods. If the study shows that the two assays are completely comparable across the measuring range (good correlation and no bias), then the reference interval can be adopted unchanged.
- Alternatively, if the study shows good correlation but a proportional negative or positive bias between the two methods, it may be acceptable to use the regression equation generated by the study to "correct" the reference interval to take account of this systematic bias.
- The guidelines provide the following example of the way this is applied:

The results of a comparison study of methods x (old method) and y (new method to be adopted) across a concentration range of 50-250 give the best-fit linear regression line:

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y = 1.57x - 0.832 correlation coefficient r2 = 0.990
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The established reference interval for method x is 50-150.

Since there is excellent correlation but proportional bias between the two methods, the "corrected" reference interval for method y can be calculated thus:

```
For the lower limit 50 y = (1.57 \times 50) - 0.832 = 77.72 (which rounds up to 78)
For the high limit 150 y = (1.57 \times 150) - 0.832 = 234.82 (which rounds up to 235)
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The reference interval to be adopted for the new method y is 78-235.

Using this method:

- A minimum of 40 patient samples should be tested across the reportable range (absence of disease, and with disease)
- Advantage of the transferring protocol is that it does not require analysis of samples from reference individuals.
- Limited application because it only applies if the reference interval in question has been in use at that particular institution.
- Most important is making the decision about whether or not the two methods agree sufficiently for them to share the same reference interval
- If that is hard to determine guidelines suggest that validation of the reference interval is indicated.

Verify An established reference interval

- **Verification** of an established reference interval is used when a laboratory adopts an established reference interval that is from a manufacturer or another laboratory using the same or similar analytical system.
- The **preanalytical protocol** for processing patient samples should not be significantly different from that used for determining reference values when establishing the reference interval.
- The **verification study** is designed to confirm that the established reference interval is **appropriate for the population** served by the adopting laboratory.
- Reference values for at least 20 healthy individuals representative of the adopting laboratory's healthy population.
- The exclusion criteria for the selection of reference individuals used should be the same as the original study.
- If the review is considered compatible with the testing laboratory, the RI can be used
- The review needs to be purposeful and documented

Select the reference individuals (n = 20) representative of the laboratory's healthy population

Determine the reference values

Check the homogeneity of these groups (no outliers)

If no more than 2 of the 20 tested subjects values fall outside the original limits

Reference limits of the donor are considered as valid

If 3 or 4 test results fall outside these limits, another 20 reference specimens should be obtained

If no more than 2 of these results fall outside the donor's RI:

the latter is considered as acceptable

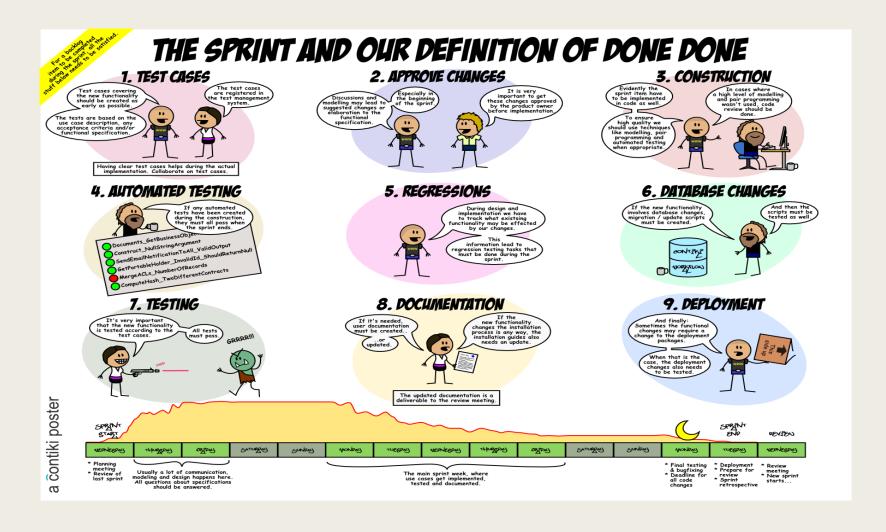
If 3 or more fall outside the RI:

- Revise the analytical procedure
- Consider possible biological differences
- Develop own RIs according to the original method

Issues with conducting a Reference Interval:

- Testing is expensive
- Procedures are labor intensive, time consuming
- Is a reference interval required, or does there need to be a cutoff
- Where do I get the subjects?

Plan, plan, plan the study



Pre-Analytical: 50-70% of errors occur here!

- Have a well written SOP that includes relevant stakeholders including phlebotomy, time frames and where and when the study will be conducted.
- Will this be a verification or validation? How many samples will be required? How much volume is required? Will additional volume be required for freezing?
- What is your institutional policy? Is there IRB approval? Do you have a consent form?
- Do you have a questionnaire, will the study be a priori or a posteri?
- Who will recruit/ consent/ draw the subjects?
- Do I have sufficient representation of males/females as well as ethnic groups that represent your testing population?
- What is exclusion criteria? Is this for only adults? Children? Geriatric?

Analytical:

- Do you have sufficient reagents to conduct the study? How many sites? How many analyzers?
- Are the analyzers working correctly? Should there be a preventative maintenance?
- Are the centrifuges working correctly?
- Will the study be conducted over a period of time to introduce analytical variables that are seen in patient testing? How many samples/day?
- Will all samples be run fresh? Will some be run frozen? Who will aliquot samples to be frozen?

Post Analytical

- What is considered an outlier?
- Who will do the statistical analysis? What type will be based on the amount of samples? Parametric, non-parametric?
- Analyze the reference values: select a statistical method and calculate the limits of reference and the reference interval
- Report sign off
- Update LIS and downstream systems
- Alert clinicians to range changes.

RI requirements

- RI should reflect patient population testing
- If possible a full validation should be conducted (120)
- If not possible, the level of uncertainty the laboratory is willing to accept needs to be considered.
- Method for determining outliers

Limitations of conducting a Reference Interval

- Reference samples are not always be possible to collect, as a diverse set of healthy individuals may not volunteer for unnecessary clinical procedures
- There is also no general agreement on how to define healthy individuals, and labs often use samples that are "easily collected," such as college students in academic clinics or internal laboratory staff.
- Some laboratories do not collect their own samples, but rather rely on reference intervals from the literature or equipment manufacturers
- Distributions of normal lab test values can vary between self-identified races and ethnicities (SIRE). To account for these differences, some test results use SIRE-specific reference intervals
- What about special samples: CSF, Amniotic Fluids?

Conclusion

- Conducting a Reference Range Interval is a complicated process
- Carefully planned and documented event
- The more defined your population and subject selection and the more controlled your pre-analytical variables are the better the outcomes
- If a RI is to be transference, should be compatible for reagent/instrument combination and the testing population
- Ensure best possible results to provide tools for clinicians to diagnose disorders and provide optimal treatment to patients